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A mathematical modeling approach to assess biological control of an orange tree disease

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Abstract

The model presented and investigated here describes the interaction between the orange tree and two different microorganisms, the pathogen fungus *Guignardia citricarpa* and the antagonist *Trichoderma harzianum* T1A. The pathogen-free and coexistence are the only possible system's equilibria. The pathogen-free points bifurcates from coexistence when the antagonist strength is sufficiently high, but does not appear to much be dependent from the amount of beneficial fungus employed. This **result** represents a relevant guideline for the applied ecologist and for the farmers. Sensitivity analysis in suitable parameter spaces is performed numerically.

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Orange tree, *Guignardia citricarpa*, *Trichoderma harzianum* T1A, mathematical model

1. Introduction

Citrus Black Spot, caused by the fungus *Phylosticta citricarpa* (previously known as *Guignardia citricarpa*), is one of the most damaging fungal citrus diseases causing significant yield losses in countries like Brazil, Australia and South Africa [23]. Citrus Black Spot is a foliar and fruit disease, characterised by hard spots, virulent spots, false melanose, freckle spots on fruit, and necrotic lesions on leaves and twigs [12]. It affects all commercial citrus cultivars, with injured fruits being commercially undeserved *in natura*, intended only for juice production.

Citrus Black Spot disease was reported for the first time in Australia [3], in 1895, and is currently present in warm, summer rainfall areas of Asia, Africa, South America and North America. Although Citrus Black Spot has never been reported in Europe, the fungus has recently been reported in several European countries (Italy, Malta and Portugal) [11], and is considered to have potential for establishment and spread [5].

Citrus fruits with Citrus Black Spot lesions are subject to quarantine regulations in the European Union [6] and USA [25]. The regulations restrict market access where fruit is quarantined; the fruit cannot be sold in the fresh market [14] thereby reducing the availability of citrus fruits to consumers in the off-season in Europe [1].

The control of Citrus Black Spot is exerted by applying fungicides, and the fruits need to be sprayed four to five times during the infection period [24]. In severe attacks, *P. citricarpa* can cause premature fruit fall, reducing plant productivity, with losses up to 80% in productivity. Fungicides have also been associated with negative environmental impacts [9] and cause the selection of strains resistant to the active principles used [10]. Biological control agents (BCA) have received large attention, and their use has contributed to the control of fungal diseases both as a complement or replacement of agrochemicals, with consequently lower ecological and economical costs. Biological control of pathogens relying on the use of living organisms to keep in check pests has been used for centuries [21]. *Trichoderma* species have been used with success as biocontrol agents [19] against numerous pathogens

33 including *P. citricarpa* [16, 17, 15, 8, 4]. *Trichoderma harzianum* T1A se-
 34 cretome inhibits the growth and mycelial melanization of *P. citricarpa*. The
 35 biocontrol agent secretes proteins related to the control of *P. citricarpa*, and
 36 induction of plant resistance, even in the absence of pathogen challenge [17].

37 The objective of this investigation is to explore and evaluate the effec-
 38 tiveness of such biological practice through a mathematical model. We de-
 39 scribe the behaviour of a biological system composed by an orange tree, the
 40 pathogen fungus, *Guignardia citricarpa*, and the beneficial one, *Trichoderma*
 41 *harzianum* T1A. In the next Section we formulate the model, which is qual-
 42 itatively analysed in Section 3. Bifurcations are investigated next. Some
 43 numerical simulations support the theoretical findings and indicate possible
 44 management strategies. The results are discussed in Section 6.

45 2. The model

46 The three populations of interest here are the orange tree fruits, O , the
 47 pathogen fungus *G. citricarpa*, P , and the beneficial fungus *T. harzianum*
 48 *T1A*, F . The model reads:

$$\begin{aligned} \frac{dO}{dt} &= rO \left(1 - \frac{O}{K}\right) - h(F)OP, \\ \frac{dP}{dt} &= eh(F)OP - cP^2 - aFP, \\ \frac{dF}{dt} &= sF \left(1 - \frac{F}{H}\right) + baFP. \end{aligned} \quad h(F) = \frac{1}{q + F}, \quad (1)$$

49 The first equation describes the evolution of the orange tree, growing
 50 logistically with net growth rate r and carrying capacity K , being negatively
 51 affected by the pathogen fungus. This however is somewhat reduced by the
 52 presence of F and modeled via the dependence on the interaction coefficient
 53 on F . The larger the amount of F , the most effective the reduction is, so
 54 that we take the variable parameter $h(F)$ to be a decreasing function.

55 The second equation contains the evolution of the pathogen fungus feed-
 56 ing on the tree parts, where $e < 1$ represents a conversion factor. Intraspecific
 57 competition for resources at rate c is taken into account. The fungus cell walls
 58 are degraded by the extracellular enzymes produced by *Trichoderma* at rate
 59 a for which *G. citricarpa* experiences a loss.

60 In the third equation the beneficial fungus feeds on resources that are not
 61 explicitly modeled, logistic term with net reproduction rate s and carrying

62 capacity H , but also obtains additional food by degrading the bad fungi with
 63 conversion coefficient $b < 1$.

64 3. The qualitative analysis of the model

65 3.1. Boundedness

66 In order to have a well-posed model, we show that the system's tra-
 67 jectories remain confined within a compact set. Consider the total system
 68 population $\varphi(t) := O(t) + P(t) + F(t)$. Summing up the equations of (1) for
 69 an arbitrary $\eta > 0$, dropping the negative terms, since $e < 1$ and $b < 1$, we
 70 then have

$$\begin{aligned} \frac{d\varphi(t)}{dt} + \eta\varphi(t) &= \Pi(O) + \chi(P) + \Gamma(F) \leq \Pi_M(O) + \chi_M(P) + \Gamma_M(F) \\ \Pi(O) &= (r + \eta)O - \frac{r}{K}O^2, \quad \chi(P) = (\eta - cP)P, \quad \Gamma(F) = (s + \eta)F - \frac{s}{H}F^2 \\ \Pi_M &= \Pi(O_M) = \frac{K(r + \eta)^2}{4r}, \quad O_M = \frac{r + \eta}{2r}, \\ \chi_M &= \chi(P_M) = \frac{\eta^2}{4c}, \quad P_M = \frac{\eta}{2c}, \\ \Gamma_M &= \Gamma(F_M) = \frac{H(s + \eta)^2}{4s}, \quad F_M = \frac{s + \eta}{2s}. \end{aligned}$$

Setting $M = \min\{\Pi_M, \chi_M, \Gamma_M\}$ and dropping the negative terms, we have
 the differential inequality $\varphi'(t) \leq M\varphi(t)$ from which, upon its solution, the
 upper bound follows

$$\varphi(t) \leq \max\{M, \varphi(0)\}.$$

71 Since each population is nonnegative, it is bounded by the same upper bound
 72 as well.

73 3.2. System's equilibria

74 System (1) has seven possible equilibria, the configuration $E_2 = (0, P_2, 0)$
 75 not being allowed because, biologically, P is a specialist predator on the
 76 orange tree and in its absence, it cannot thrive. Also $E_6 = (0, P_6, F_6)$ is
 77 unfeasible, as it is easily checked that $F_6 < 0$.

78 For stability assessment, the Jacobian of (1) is needed:

$$J = \begin{bmatrix} r - \frac{2rO}{K} - \frac{P}{q+F} & -\frac{O}{q+F} & \frac{OP}{(q+F)^2} \\ \frac{eP}{q+F} & \frac{eO}{q+F} - 2cP - aF & -\frac{eOP}{(q+F)^2} - aP \\ 0 & baF & baP + s - \frac{2sF}{H} \end{bmatrix}. \quad (2)$$

Equilibria $E_0 = (0, 0, 0)$, $E_1 = (K, 0, 0)$ and $E_3 = (0, 0, H)$, all unconditionally feasible, are all unstable. Indeed for E_0 the eigenvalues are r , 0 and s ; for E_1 they are $-r$, eKq^{-1} and s ; for E_3 we find r , $-aH$, $-s$. Further, $E_5 = (O_5, P_5, 0)$ with

$$O_5 = \frac{rq^2K}{rq^2 + ceK}, \quad P_5 = \frac{ce}{q}O_5$$

79 is also always feasible but unstable by the positive eigenvalue $s + abP_5$.

80 For the pathogen fungus-free equilibrium $E_4 = (K, 0, H)$, E_4 finally the
81 eigenvalues are $\lambda_1 = -r$, $\lambda_2 = [eK - aH(q + H)](q + H)^{-1}$ and $\lambda_3 = -s$
82 giving the stability condition

$$eK < aH(q + H). \quad (3)$$

83 For coexistence, $E_* = (O_*, P_*, F_*)$, solving the third equation of (1) and
84 substituting it into the first one, we get

$$P_* = \frac{sF_* - Hs}{Hba}, \quad O_* = \frac{HKbarq + HKs + F_*(HKbar - Ks)}{Hbar(rq + eF_*)} \quad (4)$$

85 where F_* is a real positive root of the cubic $\Psi(F) := AF^3 + BF^2 + CF + D = 0$
86 with $A = (Ha^2b + cs)r > 0$, $D = -H[esK + qr(Kabe + cqs)] < 0$, which
87 ensure at least one positive root, and

$$B = [((a^2bq - cs)H + cqs + q(Ha^2b + cs)]r \quad (5)$$

$$C = [((a^2bq - cs)H + cqs)q - H(Kabe + cqs)]r - sKe.$$

88 However, nonnegativity of O_* and P_* must be ensured. It follows from

$$\frac{s - Hbar}{s + barq} < \frac{H}{F_*} < 1. \quad (6)$$

The Routh-Hurwitz stability conditions in this case are

$$-\text{tr}(J(E_*)) = \frac{r}{K}O_* + cP_* + sF_* > 0,$$

89 which holds, and letting $M^{(2)}$ be the sum of the minors of order two of $J(E_*)$,

$$M^{(2)} = \frac{r}{K}O_*P_* + \frac{eP_*O_*}{(q+F_*)^2} + s\frac{r}{K}F_*O_* + csF_*P_* + abF_* \left(a + \frac{eO_*}{(q+F_*)^2} \right) P_*,$$

$$\det(J(E_*)) = F_*O_*P_* \left[\frac{crs}{K} + \frac{abeP_*}{(q+F_*)^3} - \frac{abr}{K} \left(a + \frac{eO_*}{(q+F_*)^2} \right) - \frac{es}{(q+F_*)^2} \right],$$

90 so that the remaining stability conditions are given by

$$\det(J(E_*)) < 0, \quad \text{tr}(J(E_*))M^{(2)} < \det(J(E_*)), \quad (7)$$

91 which we avoid to write down explicitly.

92 Table 1 summarizes feasibility and stability of the system's equilibria.

Table 1: Characterization of the equilibria of (1).

$E = (O, P, F)$	Feasibility conditions	Stability conditions
$E_0 = (0, 0, 0)$	—	unstable
$E_1 = (K, 0, 0)$	—	unstable
$E_2 = (0, P, 0)$	unfeasible	—
$E_3 = (0, 0, H)$	—	unstable
$E_4 = (K, 0, H)$	—	(3)
$E_5 = (O_5, P_5, 0)$	—	unstable
$E_6 = (0, P_6, F_6)$	unfeasible	—
$E_* = (O_*, P_*, F_*)$	(6)	(7)

93 4. Bifurcation analysis

94 Concentrating on the pathogenic fungus-free equilibrium, we observe that
 95 the eigenvalue λ_2 vanishes for the parameter choice

$$a^\dagger = \frac{eK}{H(q+H)}. \quad (8)$$

The Jacobian evaluated at E_4 simplifies to

$$J = \begin{bmatrix} -r & -\frac{K}{q+H} & 0 \\ 0 & 0 & 0 \\ 0 & abH & -s \end{bmatrix}$$

96 for which the right and left eigenvectors respectively are $\mathbf{v} = (v_1, 1, v_3)^T$,
 97 with $v_1 = -K[r(q+H)]^{-1}$, $v_3 = abHs^{-1}$, and $\mathbf{w} = (0, 1, 0)^T$. Also, denoting
 98 by $\mathbf{f}(O, P, F) = (\mathbf{f}^1, \mathbf{f}^2, \mathbf{f}^3)$ the right hand side of (1), and by subscripts the
 99 partial derivatives, we find

$$\mathbf{f}_a = \begin{pmatrix} 0 \\ -PF \\ bPF \end{pmatrix} \quad D\mathbf{f}_a = \begin{bmatrix} 0 & 0 & 0 \\ 0 & -F & -P \\ 0 & bF & bP \end{bmatrix}, \quad (9)$$

from which $\mathbf{f}_a(E_4, a^\dagger) = \mathbf{0}$ and consequently $\mathbf{w}^T \mathbf{f}_a(E_4, a^\dagger) = 0$. This result
 represents the first condition in Sotomayor's theorem, [22], that we use at
 present. Further, $\mathbf{w}^T D\mathbf{f}_a(E_4, a^\dagger) \mathbf{v} = -H \neq 0$, the second required condition
 for the existence of bifurcations. We then need to evaluate

$$\mathbf{w}^T D^2 \mathbf{f}(E_4, a^\dagger)(\mathbf{v}, \mathbf{v}) = D^2 \mathbf{f}^2(E_4, a^\dagger)(\mathbf{v}, \mathbf{v}).$$

100 Observe that the second partial derivatives of \mathbf{f}^2 are $\mathbf{f}_{OO}^2 = 0$, $\mathbf{f}_{PP}^2 = -2c$ and
 $\mathbf{f}_{OP}^2 = \frac{e}{q+F}$, $\mathbf{f}_{OF}^2 = -\frac{eP}{(q+F)^2}$, $\mathbf{f}_{PF}^2 = -a - \frac{eO}{(q+F)^2}$, $\mathbf{f}_{FF}^2 = \frac{2eOP}{(q+F)^3}$

101 and thus at this equilibrium all vanish, except three. In summary, we find

$$\begin{aligned} D^2 \mathbf{f}^2(E_4, a^\dagger)(\mathbf{v}, \mathbf{v}) &= 2 \frac{e}{q+F_4} v_1 - 2c - 2 \left(a^\dagger + \frac{eO_4}{(q+F_4)^2} \right) v_3 \\ &= -2 \frac{eK}{r(q+H)^2} - 2c - 2 \frac{a^\dagger bH}{s} \left[a^\dagger + \frac{eK}{(q+H)^2} \right] \\ &= -2eK \left[\frac{1}{r(q+H)^2} + \frac{c}{eK} + \frac{q+2H}{(q+H)^3} \frac{beK}{sH} \right] \neq 0. \end{aligned}$$

102 so that also the third condition for the occurrence of a transcritical bifur-
 103 cation is satisfied. Since in this case when a crosses from above the critical
 104 threshold a^\dagger the equilibrium E_4 becomes unstable and the P population ap-
 105 pears in the system, the latter settles to the coexistence equilibrium and the
 106 pathogen establishes permanently in the system.

107 **5. Numerical simulations and discussion**

108 The findings of the previous section are also supported by numerical ex-
 109 periments. Figure 1 contains the bifurcation diagram of O , P , F in terms of
 110 the degradation rate a of P by F .

- 111 (a) For $a = 0.1$ a transcritical bifurcation arises between the coexistence
 112 equilibrium and the pathogen fungus-free equilibrium, E_4 .
- 113 (b) For $a < 0.1$ an interesting behaviour occurs. Increasing a increments
 114 the density of the beneficial fungus but when the degradation rate
 115 a attains a threshold, here 0.05, the beneficial fungus experiences a
 116 decline in the reward gotten from the action of the pathogen fungus,
 117 because the density of the latter becomes too low.
- 118 (c) For $a > 0.1$ the system (1) attains the pathogen-free point, the most
 119 important ecological equilibrium, with both O and F at their respective
 120 carrying capacities K and H . Note that the equilibrium populations
 121 at E_4 do not depend on a , that no change in their values occurs by
 122 increasing a .

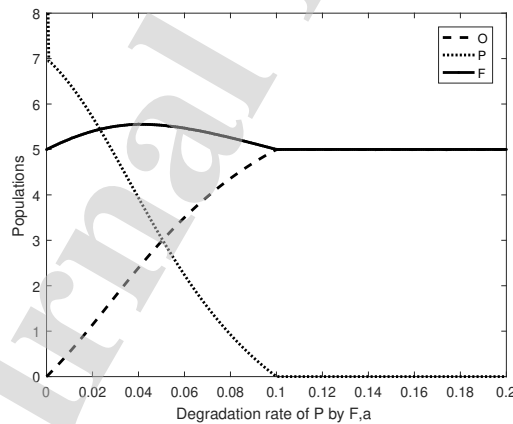


Figure 1: The bifurcation diagram of O (dashed), P (dotted) and F (continuous line) with respect to the degradation rate of P by F , a . The other parameters values are $r = 1$, $K = 5$, $e = 0.7$, $b = 0.7$, $s = 1$, $H = 5$, $q = 2$ and the i.c. are $(1, 1, 1)$. With these parameter values it can easily be checked that $a^\dagger = 0.1$ coinciding with the value expected from (8), so that a transcritical bifurcation arises between E^* and E_4 .

123 Figure 2 shows the densities of the populations O , P and F , at steady
 124 state, in terms of two model parameters: (a, e) in panel (a); (a, b) in panel
 125 (b); (a, q) in panel (c). In all these situations by fixing the value of the
 126 parameter on the y -axis and increasing a leads to an increase in the density
 127 of O , in an decrease in the density of P , while for F at the beginning there is
 128 an increase followed by a decrease. This latter feature corresponds to what is
 129 observed in Figure 1. Let us now fix the value of a . In panel (a) we find that
 130 increasing the parameter e , both P and F increase, while O decreases. In
 131 panel (b) increasing b , O experiences a slow increase, P has a slow decrease
 132 and F increases. In panel (c) increasing q , both O and P increase while F
 133 first increases and then decreases.

134 The biological control of Citrus Black Spot has been shown but only
 135 by scarce studies. Among them, the use of *Saccharomyces cerevisiae* Meyer
 136 [20, 7, 8] and the possibility of using volatile organic compounds produced
 137 by yeast was reported. Also, bacteria of the genus *Bacillus*, such as *B.*
 138 *thuringiensis* var. *kurstaki* (HD-1), obtained from the commercial products
 139 Dipel®WP and Dimy Pel®, and *B. thuringiensis* var. *kurstaki* (HD-567),
 140 used in the control of *P. citricarpa*, demonstrated to reduce the number of
 141 picnids per *P. citricarpa* lesion, and the number of lesions per fruit [18].

142 Guimarães, [13] used as an antagonist the fungus *Trichoderma koningii*
 143 isolated from the surface of “Montenegrina” tangerine leaves. The authors
 144 were able to control *P. citricarpa* in vitro and in vivo using the same orchard
 145 from where the antagonist was isolated. It has also been demonstrated that
 146 *T. koningii* reduces the severity and incidence of Citrus Black Spot.

147 Other organisms or combinations of microorganisms may have different
 148 efficiencies when compared to experiments under field conditions. [4] used *T.*
 149 *harzianum*, *B. subtilis* and a biofertilizer, which had a microbial load com-
 150 posed mainly of bacteria (*Bacillus* spp, *Pseudomonas* spp and actinobacteria)
 151 to control *P. citricarpa* in citrus fruits. Best results were achieved with the
 152 biofertilizer, followed by *B. subtilis* and *Trichoderma*. As pointed out by [15],
 153 antagonism of fungal pathogens under laboratory conditions may not be re-
 154 flected under field conditions. The authors observed that *Bacillus subtilis*
 155 was able to control *P. citricarpa* in vitro, but the efficiency under field was
 156 not reproducible, requiring further studies to select more efficient isolates
 157 and the best application period.

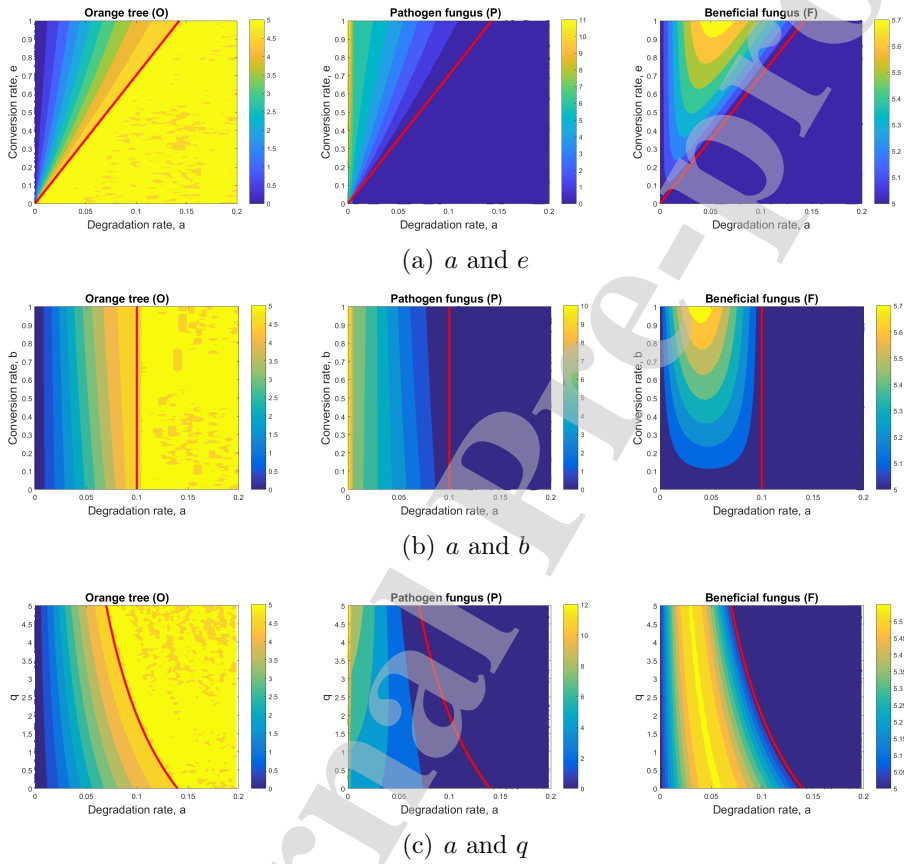


Figure 2: The density of O , P and F at stable state, as function of two parameters. The red curve, obtained by (3), partitions the domain into the stable coexistence equilibrium (on its left) and the pathogen-free point E_4 (on its right). Note that the axes of the colorbars are different in each figure.

158 6. Conclusions

159 Recent studies [16, 17] show that *T. harzianum* T1A and *Trichoderma*
 160 *atroviride* T17 are able to control *Guignardia citricarpa* Gc3. Unlike other
 161 *T. harzianum* strains, T1A is able to induce plant resistance without being
 162 challenged by a pathogen.

163 Mathematical analysis is instrumental in defining strategies for the biolog-
 164 ical control of this infestant. Policies to possibly use *Trichoderma harzianum*
 165 *T1A* in orange cultures to fight *Guignardia citricarpa* and reduce farmers'
 166 economic losses can be devised on this analysis.

167 The system has two possible equilibria, the pathogen fungus-free point
 168 and coexistence. Condition (8) states that from the latter the system set-
 169 tles to the former if the degradation rate of fungus cell walls a by *Tricho-*
 170 *derma harzianum T1A* falls below the critical threshold a^\dagger . In such case
 171 the pathogen *Guignardia citricarpa* becomes endemic. However, note that
 172 annihilating λ_2 for the pathogen-free equilibrium E_4 can be obtained also
 173 by parameter choices other than a , (8). For instance, one can choose $e^\dagger =$
 174 $aH(q + H)K^{-1}$. In this case it is easily checked that both $\mathbf{w}^T \mathbf{f}_e(E_4, e^\dagger) = 0$
 175 and $\mathbf{w}^T D\mathbf{f}_e(E_4, e^\dagger)\mathbf{v} = K(1 + H)^{-1} \neq 0$ still hold. As no change in the second
 176 derivatives $D^2\mathbf{f}$ occurs, the transcritical bifurcation arises also crossing the
 177 critical value e^\dagger . From an ecological point of view e is a conversion factor,
 178 that measures the efficiency on how the pathogen fungus is able to exploit
 179 the nutrients obtained from the orange fruits. When its value is 1, maximum
 180 efficiency is obtained. But, independently of the value, if e falls below the
 181 critical value e^\dagger , E_4 , the pathogenic fungus-free point is stable, while if it lies
 182 above the threshold e^\dagger , E_4 becomes unstable. The system trajectories then
 183 move to the only other possible allowed stable equilibrium, E_* . This change
 184 in the ultimate behavior of the system represents the meaning of a transcriti-
 185 cal bifurcation. The coexistence equilibrium E_* contains all the populations,
 186 and therefore the pathogen *Guignardia citricarpa* invades the ecosystem and
 187 becomes endemic in it. To remove the pathogenic fungus or keep the system
 188 pathogen-free, the conversion factor should be less than the threshold, $e < e^\dagger$.
 189 Because the conversion factor is an intrinsic property of the fungus, it is hard
 190 to be altered by human actions. However, the value of the threshold can be
 191 raised, to enhance the satisfaction of the inequality. This could be achieved
 192 if the carrying capacity K of the orange tree is lowered, or conversely the
 193 carrying capacity H of the beneficial fungus is raised. These measures can
 194 likely be more easily implemented by the farmers, especially for instance by

195 pruning the orange tree. Note that the same measures have the opposite ef-
 196 fect on the degradation rate's a critical threshold a^\dagger , of the fungus cell walls,
 197 lowering it, but with the ultimate same effect on the ecosystem.

198 Instead, attempting to use H as bifurcation parameter, for which the crit-
 199 ical value would be obtained by the positive root of the quadratic $aH^2 + aqH -$
 200 $eK = 0$, namely $H^\dagger = (2a)^{-1}[\sqrt{a^2q^2 + 4aeK} - aq]$, leads to $\mathbf{w}^T \mathbf{f}_H(E_4, H^\dagger) =$
 201 0 as well as $\mathbf{w}^T D\mathbf{f}_H(E_4, H^\dagger)\mathbf{v} = 0$, for which no bifurcation can arise. Sim-
 202 ilarly, taking $K^\dagger = aH(q + H)e^{-1}$, it follows $\mathbf{w}^T \mathbf{f}_K(E_4, K^\dagger) = 0$ as well
 203 as $\mathbf{w}^T D\mathbf{f}_K(E_4, K^\dagger)\mathbf{v} = 0$ and again no bifurcation can arise. **This entails**
 204 **that the measures discussed above on pruning the trees, thus lowering K ,**
 205 **or fostering the *Trichoderma harzianum* T1A, cannot alone directly lead to**
 206 **a transcritical bifurcation and consequent *Guignardia citricarpa* eradication.**
 207 **Nevertheless, as they lead to favorable changes in the thresholds e^\dagger and a^\dagger ,**
 208 **their exploitation combined with perhaps small modifications either in the**
 209 **conversion factor e or the degradation rate a could help for the pathogen**
 210 **eradication.**

211 **We can conclude that** these remarks are important for the ecologist work-
 212 ing in the field, and constitute a guideline for possible economic advantages
 213 for the farmer. Indeed, they hint that to eradicate the pest, the effectiveness
 214 of the antagonist here expressed by the parameters a and e , has more influ-
 215 ence than the amount that is actually sprayed, represented in a sense by the
 216 fungus carrying capacity H , or alternatively by the size of the tree receiving
 217 the treatment, modeled via its "size" K .

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222 **Conflict of interest** The authors declare that they have no conflict of
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