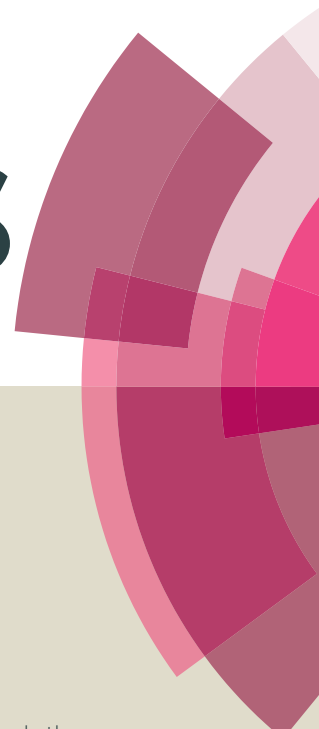


# RSC Advances



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1 **A Robust Fluorescent Chemosensor for Aluminium Ion Detection**  
2 **Based on Schiff base Ligand with Azo Arm and Application in**  
3 **Molecular Logic Gate**

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9  
10 **Abstract**

11 In this present work we reported synthesis and structural characterisations of a N2O2  
12 donor Schiff base chemosensor with azo arm (**H<sub>2</sub>L**). Various spectroscopic tools like single  
13 crystal X-ray, NMR, UV-vis, FTIR, ESI-Mass etc have been deployed to develop the present  
14 work. In recent past years a number of azo derived chemosensors have been reported by different  
15 research groups. This is first time we are reporting designing and properties of an azo derived  
16 chemosensors (**H<sub>2</sub>L**) for detection of aluminium ion in semi aqueous medium. It has been found  
17 that it selectively senses Al<sup>3+</sup> ion in semi aqueous solution. Here, the sensing process is mainly  
18 based on Chelation enhanced fluorescence process (CHEF). It has very high selectivity over  
19 other metal ions and anions. A detailed literature survey has been carried out and compared with  
20 this work. It has appreciably low detection limit i.e. 6.93 nM.<sup>1</sup>HNMR titration was done to  
21 support the plausible complexation process. 1:1 stoichiometry binding between the chemosensor  
22 and Al<sup>3+</sup> ion has been confirmed from Job's plot. An inhibition molecular logic gate has been  
23 constructed using Chemosensor (**H<sub>2</sub>L**), where Al<sup>3+</sup> and EDTA act as inputs and Fluorescence

24 emission as output. The structural and electronic parameters of the chemosensor ( $H_2L$ ) and  
25 complex  $[AL(L)]NO_3$  have been studied in details using theoretical tools like DFT and TDDFT.

26

## 27 **Introduction**

28 The design and construction of powerful and highly selective fluorescent chemosensors have  
29 attracting considerable attention in biomimetic chemistry.<sup>1</sup> Among various heavy metal ions  
30 Aluminium is an essential element due to its abundance and use in every sphere of life as  
31 utensils, electronic and electrical components of different gadgets, building equipment, different  
32 packaging items, water treatments, food additives, pharmaceutical products, occupational dusts  
33 etc. Aluminum is a known neurotoxin to organisms<sup>1</sup> and is believed to cause Alzheimer's  
34 disease,<sup>2</sup> osteomalacia<sup>3</sup> and breast cancer.<sup>4</sup> Toxicity due to the presence of excess aluminium in  
35 human health may arise as it inhibits several essential elements of similar size and charge like  
36  $Mg^{2+}$ ,  $Ca^{2+}$  and  $Fe^{3+}$ . It is also responsible for retarded growth of plants<sup>5</sup> and oxidative damage of  
37 the cell membrane.<sup>6</sup> In spite of its many drawbacks, the enormous use of Al in daily life causes  
38 accumulation of  $Al^{3+}$  and hence toxicity towards human health and the environment. WHO  
39 recommended the average weekly human body dietary intake of  $Al^{3+}$  of around 7 mg/kg body  
40 weight.<sup>7</sup> Thus, it is utmost urgent to detect  $Al^{3+}$  ion in environment in trace level i.e. in ppb, ppm  
41 or nano level. Unfortunately, the determination of  $Al^{3+}$  is complicated mainly due to its poor  
42 coordination ability, strong tendency to hydration, and lack of suitable spectroscopic  
43 characteristics.<sup>8</sup> Some analytical methods are available for detection of  $Al^{3+}$  such as graphite  
44 furnace atomic absorption spectrometry, inductively coupled plasma atomic emission  
45 spectrometry, atomic absorption spectra,<sup>9</sup> electrochemistry,<sup>10</sup> mass spectrometry,<sup>11</sup> and  $^{27}Al$   
46 NMR technology.<sup>12</sup> Most of them are expensive and time consuming. Optical detection

47 particularly fluorescence spectroscopic technique is advantageous over the other techniques  
48 because of its operational simplicity, low cost, low detection limit, real-time detection,  
49 selectivity, time saving and environmental friendly property. Therefore design and synthesis of  
50 new chemosensor for the detection of  $\text{Al}^{3+}$  has received considerable attention. Recently various  
51 research groups have reported different organic probes which can selectively detect  $\text{Al}^{3+}$ .<sup>13,14</sup>  
52 Although example of such  $\text{Al}^{3+}$  detecting chemosensors are still less compared with other metal  
53 ions detecting chemosensors like  $\text{Zn}^{2+}$  mainly due to less coordinating ability of  $\text{Al}^{3+}$  compared  
54 to other metal ions. Therefore, more improvement in this field is still required. Current literature  
55 survey (Table S1) reveals that preparation of most of the chemosensors for detection of  $\text{Al}^{3+}$   
56 involves expensive starting materials, multiple reaction steps, use of different solvent mixtures  
57 and maintenance of drastic reaction conditions i.e. very high or low temperature, presence of  
58 catalyst etc. G. Das and *et al.* in a recent work<sup>14n</sup> have reported an  $\text{Al}^{3+}$  sensing organic probe viz.  
59 (E)-N'-((E)-3-(4-(dimethylamino)phenyl)allylidene)picolinohydrazide using starting materials  
60 picolinohydrazide and 4-(dimethylamino)cinnamaldehyde. Preparation of picolinohydrazide  
61 involves multiple steps. Preparation of another chemosensor viz. 2-((naphthalen-6-  
62 yl)methylthio)ethanol involves inert atmosphere and finally the product has been purified  
63 through column chromatography with 65% yield.<sup>14e</sup> S. Goswami and coworkers have reported a  
64 spirobenzopyran-quinoline (SBPQ) based sensor which selectively detects  $\text{Al}^{3+}$  along with  $\text{Fe}^{3+}$   
65 and  $\text{Cr}^{3+}$ . Preparation of this type of chemosensors also involves multiple steps.<sup>14o</sup> It is also  
66 important to mention that 2-hydroxynaphthaldehyde based organic probes are mostly used for  
67 fluorescence sensing of  $\text{Al}^{3+}$ .<sup>14p</sup>

68 Different mechanistic pathway like intramolecular charge transfer (ICT),<sup>15</sup> photo induced  
69 electron transfer (PET),<sup>16</sup> chelation-enhanced fluorescence (CHEF),<sup>17</sup> metal–ligand charge

70 transfer (MLCT),<sup>18</sup> excimer/exciple formation, imine isomerization,<sup>19</sup> intermolecular hydrogen  
71 bonding,<sup>20</sup> excited-state intramolecular proton transfer,<sup>21</sup> displacement approach,<sup>22</sup> and  
72 fluorescence resonance energy transfer<sup>23</sup> are adopted to explain the action of different  
73 fluorescence probe. Among different types of chemosensors, Schiff bases incorporated with  
74 different functionalized moiety are widely used mainly due to their ease of synthesis and low  
75 cost.

76 In this work an azo based salen-type of Schiff base ligand (E)-6,6'-((1E,1'E)-(propane-  
77 1,3-diylbis(azanylylidene))bis(methanylylidene))bis(2-methoxy-4-((E)-phenyldiazenyl)phenol)  
78 (**H<sub>2</sub>L**) is structurally characterized and it exhibit high selectivity towards Al<sup>3+</sup> ( $3.31 \times 10^3 \text{ M}^{-1}$ )  
79 with low detection limit (6.93 nM). Azo derivatives have prevalent use in different fields like  
80 pharmaceuticals, optical data storage, non-linear optics, photo switching devices, textile industry  
81 dye-sensitized solar cells and they also act as fluorescent chemosensors.<sup>24-30</sup> Our work is  
82 important and novel in this aspect because although various azo containing chemosensors are  
83 reported in literature but to the best of our knowledge in this work we are first time reporting one  
84 such sensor which can selectively detect Al<sup>3+</sup>. Our organic probe possesses some extra  
85 advantages. We have prepared **H<sub>2</sub>L** using easily available starting materials. The azoaldehyde  
86 was prepared from well known easy diazotization process with very high yield. The azoaldehyde  
87 upon reaction with a simple diamine produced **H<sub>2</sub>L** with >90% yield. We have successfully  
88 isolated both the azoaldehyde and Schiff base **H<sub>2</sub>L** in highly crystalline form. The ligand is  
89 characterized by different techniques including x-ray crystallography. The composition of the  
90 Al<sup>3+</sup>-ligand complex (Complex **1**) has been established by different spectroscopic data like IR,  
91 Mass, Job's plot and <sup>1</sup>H NMR. The DFT computation of optimized geometry of **H<sub>2</sub>L** and the  
92 complex [Al(L)]<sup>+</sup> has been used to support the electronic spectral properties.

93

## 94 **Experimental**

### 95 **Materials and Physical measurements**

96 All reagent or analytical grade chemicals and solvents were purchased from commercial  
97 sources and used without further purification. Elemental analysis for C, H and N was carried out  
98 using a Perkin–Elmer 240C elemental analyzer. Infrared spectra ( $400\text{--}4000\text{ cm}^{-1}$ ) were recorded  
99 from KBr pellets on a Nicolet Magna IR 750 series-II FTIR spectrophotometer. Absorption  
100 spectra were measured using a UV-2450 spectrophotometer (Shimadzu) with a 1-cm-path-length  
101 quartz cell. Measurements of NMR spectra were conducted using a Bruker 300 spectrometer in  
102 DMSO- $d_6$  and  $\text{CDCl}_3$  respectively. Emission was examined by LS 55 Perkin–Elmer  
103 spectrofluorimeter at room temperature (298 K) in HEPES buffer at pH= 7.4 solution under  
104 degassed condition. Fluorescence lifetimes were measured using a time-resolved  
105 spectrofluorimeter from IBH, UK.

### 106 **Synthesis of (E)-5-(2-phenyldiazenyl)-2-hydroxy-3-methoxybenzaldehyde**

107 4 mL conc. HCl was added to 15 mL water and kept in an ice bath keeping the  
108 temperature  $0^\circ\text{C}$ . To it, aniline (4.0 mmol, 0.372 g) was added. To this mixture an aqueous  
109 solution of sodium nitrite (6.0 mmol, 0.414 g) was drop wise added over a time of 30 mins. The  
110 mixture was stirred for 1 h at  $0^\circ\text{C}$ . Then the resultant solution was added to the alkaline *o*-  
111 Vanillin (4.8 mmol, 0.732 g) which is kept in another ice bath. The mixture was further stirred  
112 for 1 h at  $0^\circ\text{C}$ . The solution was neutralised using dilute HCl and  $\text{p}^{\text{H}}$  was maintained around 7.0.  
113 The mixture was extracted with chloroform and evaporated to result red coloured crystals of the  
114 aldehyde.

115 Yield: 0.953 g (94%). Anal. Calc. for  $C_{14}H_{12}N_2O_3$ : C 64.65%; H 5.79%; N 16.34%. Found: C  
116 64.36%; H 5.31%; N 16.17%. IR ( $cm^{-1}$ , KBr):  $\nu(N=N)$  1459 s;  $\nu(C-H)$  762 s. ESI-MS (positive) in  
117 MeOH: The base peak was detected at  $m/z = 279.00$ , corresponding to  $[M+23]^+$ .  
118  $^1H$  NMR ( $CDCl_3$ , 300 MHz)  $\delta$  ppm: 4.01 (-OCH<sub>3</sub>) (s, 3H), 7.25-7.35 (Ar-H) (m, 2 H), 7.74-7.86  
119 (Ar-H) (m, 5 H), 10.02 (-CH=O) (s, 1H), 11.05 (-OH) (bs, 1H).

## 120 Synthesis of Schiff base ligand (H<sub>2</sub>L)

121 A mixture of (E)-5-(2-phenyldiazenyl)-2-hydroxy-3-methoxybenzaldehyde (2.0 mmol)  
122 and 1,3-diaminopropane (1.0 mmol, 0.074 g) was heated to reflux for 4 h in methanol-  
123 chloroform solvent mixture (1:1, v/v). Upon slow evaporation of the solvent mixture deep red  
124 colored crystals were obtained.

125 Yield: 0.5282 g (96%). Anal. Calc. for  $C_{29}H_{26}N_6O_2$ : C 67.62%; H 5.49%; N 15.26%. Found: C  
126 66.86%; H 5.31%; N 14.97%. IR ( $cm^{-1}$ , KBr):  $\nu(C=N)$  1648s;  $\nu(N=N)$  1455 s;  $\nu(C-H)$  764 s. ESI-  
127 MS (positive) in MeOH: The base peak was detected at  $m/z = 551.21$ , corresponding to  $[M+1]^+$ .  
128 UV-Vis,  $\lambda_{max}$  (nm), ( $\epsilon$  ( $dm^3 mol^{-1} cm^{-1}$ )) in Acetonitrile: 290 (4533) and 388 (5736).

129  $^1H$  NMR ( $CDCl_3$ , 300 MHz)  $\delta$  ppm: 2.25 (-CH<sub>2</sub>) (m, 1H), 3.82-3.84 (-CH<sub>2</sub>) (bs, 2H), 4.01 (-  
130 OCH<sub>3</sub>) (s, 3H), 7.42-7.90 (Ar-H) (m, 7 H), 8.43 (-CH=N) (s, 1H).

131  $^1H$  NMR ( $DMSO-d_6$ , 300 MHz)  $\delta$  ppm: 2.08 (-CH<sub>2</sub>) (m, 2H), 3.74-3.82 (-CH<sub>2</sub>, -OCH<sub>3</sub>) (bs,  
132 10H), 7.30-7.48 (Ar-H) (m, 8H), 7.66-7.74 (Ar-H) (m, 6H), 8.66 (-CH=N) (s, 2H).

133  $^{13}C$  NMR ( $DMSO-d_6$ , 75 MHz)  $\delta$  ppm: 31.46 (-CH<sub>2</sub>), 56.34 (-CH<sub>2</sub>), 118.22, 122.58, 126.93,  
134 127.41, 129.08 and 130.47 (Ar-C), 165.38 (-CH=N).

135

### 136 **Synthesis of [Al(L)](NO<sub>3</sub>) complex**

137 A 2 ml methanolic solution of aluminium nitrate nonahydrate (1.0 mmol, 0.375 g) was  
138 added drop wise to 20 mL methanolic solution of **H<sub>2</sub>L** (1.0 mmol, 0.551 g) followed by addition  
139 of triethylamine (2.0 mmol, ~0.4 mL) and the resultant reaction mixture was stirred for 4 h under  
140 nitrogen atmosphere. Then the resultant mixture was dried to powdered form and further  
141 characterizations were carried out.

142 Yield: 0.6246 g (98%). Anal. Calc. for AlC<sub>29</sub>H<sub>24</sub>N<sub>7</sub>O<sub>5</sub>: C 67.62%; H 5.49%; N 15.26%. Found:  
143 C 66.86%; H 5.31%; N 14.97%. IR (cm<sup>-1</sup>, KBr):  $\nu(\text{C}=\text{N})$  1646s;  $\nu(\text{N}=\text{N})$  1545 s;  $\nu(\text{C}-\text{H})$  770 s.  
144 ESI-MS (positive) in MeOH: The base peak was detected at  $m/z = 575.25$ , corresponding to  
145 [Al(L)]<sup>+</sup>. UV-Vis,  $\lambda_{\text{max}}$  (nm), ( $\epsilon$  (dm<sup>3</sup>mol<sup>-1</sup>cm<sup>-1</sup>)) in Acetonitrile: 286 (5083) and 375 (5949).

146 <sup>1</sup>H NMR (DMSO-d<sub>6</sub>, 300 MHz)  $\delta$  ppm: 2.16 (-CH<sub>2</sub>) (bs, 2H), 3.89-3.95 (-CH<sub>2</sub>, -OCH<sub>3</sub>) (bs,  
147 10H), 7.54-7.81 (Ar-H) (m, 14H), 9.24 (-CH=N) (s, 2H).

### 148 **X-ray crystallography**

149 Single crystal X-ray data of azoaldehyde and Schiff base ligand (**H<sub>2</sub>L**) were collected on  
150 a Bruker SMART APEX-II CCD diffractometer using graphite monochromated Mo K $\alpha$   
151 radiation ( $\lambda = 0.71073$  Å) at room temperature. Data processing, structure solution, and  
152 refinement were performed using Bruker Apex-II suite program. All available reflections  $2\theta_{\text{max}}$   
153 were harvested and corrected for Lorentz and polarization factors with Bruker SAINT plus.<sup>31</sup>  
154 Reflections were then corrected for absorption, inter-frame scaling, and other systematic errors  
155 with SADABS.<sup>32</sup> The structures were solved by the direct methods and refined by means of full  
156 matrix least-square technique based on F<sup>2</sup> with SHELX-1997 and SHELX-2013 software  
157 package.<sup>33</sup> All the non-hydrogen atoms were refined with anisotropic thermal parameters. C-H



158 hydrogen atoms were inserted at geometrical positions with  $U_{\text{iso}} = 1/2U_{\text{eq}}$  to those they are  
159 attached. Crystal data and details of data collection and refinement are summarized in Table 1.

## 160 **Computational method**

161 All computations were performed using the GAUSSIAN09 (G09)<sup>33</sup> software package.  
162 Coordinates obtained from single crystal X-ray data were used for optimization of structure of  
163 Ligand (**H<sub>2</sub>L**). For optimization we used the density functional theory method at the B3LYP  
164 level<sup>35,36</sup> and the standard 6-31+G(d) basis set for C, H, N and O atoms<sup>36,38</sup> and the lanL2DZ  
165 effective potential (ECP) set of Hay and Wadt<sup>39-41</sup> for aluminum atom have been chosen for  
166 optimization .

167 TDDFT calculation was performed with the optimized geometry to ensure only positive  
168 eigen values. Time-dependent density functional theory (TDDFT)<sup>42-44</sup> was performed using  
169 conductor-like polarizable continuum model (CPCM)<sup>44-47</sup> and the same B3LYP level and basis  
170 sets in aqueous solvent system. GAUSSSUM<sup>48</sup> was used to calculate the fractional contributions  
171 of various groups to each molecular orbital.

## 172 **Results and discussion**

### 173 **Synthesis and Characterization**

174 The azoaldehyde5-(2-phenyldiazenyl)-2-hydroxy-3-methoxybenzaldehyde is synthesized  
175 by first diazotization of primary aromatic amine (aniline) followed by coupling with an aromatic  
176 alcohol (o-vanillin). Azoaldehyde reacted with 1,3-diamino propane in 2:1 molar ratio at ambient  
177 temperature in methanol to generate the Chemosensor **H<sub>2</sub>L** (Scheme 1). The yield of both the  
178 aldehyde and **H<sub>2</sub>L** is >90%. Both of them are crystallized from slow evaporation of methanol-  
179 chloroform (3:5) mixture. They are well characterized by <sup>1</sup>H NMR, <sup>13</sup>C NMR, mass and IR

180 spectroscopy. In IR spectrum, azo (N=N) band of both azoaldehyde and **H<sub>2</sub>L** appear at 1459 cm<sup>-1</sup>  
181 and 1455 cm<sup>-1</sup> respectively (Fig. S1). Other important stretching vibrations of **H<sub>2</sub>L** are 1648 (s,  
182  $\nu(\text{C}=\text{N})$ ); 3100 cm<sup>-1</sup> ( $\nu(-\text{OH})$ ); 764 cm<sup>-1</sup> ( $\nu(\text{C}-\text{H})$ ) respectively. Similar characteristic stretching  
183 frequencies of [Al(L)]NO<sub>3</sub> are obtained at 1646 (s,  $\nu(\text{C}=\text{N})$ ); 1530cm<sup>-1</sup> ( $\nu(\text{N}=\text{N})$ ); 770 cm<sup>-1</sup>( $\nu(\text{C}-$   
184 H)), respectively (Fig. S2). In complex **1**, nitrate ion is present as a counter anion. The  
185 characteristic stretching frequencies are obtained at 765 cm<sup>-1</sup> (planar rock), 815 cm<sup>-1</sup> (NO,  
186 deformation) and 1375 cm<sup>-1</sup> (NO, asymmetric stretch), respectively, which are comparable with  
187 that of previously reported literature values.<sup>49</sup> In <sup>1</sup>H NMR spectrum of azoaldehyde (CDCl<sub>3</sub>  
188 solvent), aldehyde proton (-CH=O) appeared at 10.02 ppm whereas -OCH<sub>3</sub> and phenolic -OH  
189 protons appeared at 4.01 ppm and 11.05 ppm respectively. Azoaldehyde contains two aromatic  
190 rings. Aromatic protons came as multiplet at two different regions i.e. from 7.25 to 7.35 ppm  
191 and from 7.74 to 7.86 ppm respectively. In case of the **H<sub>2</sub>L** (CDCl<sub>3</sub> solvent), aromatic protons  
192 appear around 7.74-7.30 ppm whereas azomethine proton, -OCH<sub>3</sub> protons and aliphatic protons  
193 appeared at 8.43, 4.01, 3.95 and 2.25 ppm respectively. Here, Phenolic -OH proton is so labile  
194 that it becomes very difficult to arrest in both CDCl<sub>3</sub> and DMSO-d<sub>6</sub> solvent system. In <sup>13</sup>C  
195 spectra of **H<sub>2</sub>L** (DMSO-d<sub>6</sub> solvent), aliphatic carbon signals appeared at 31.46 ppm and 56.34  
196 ppm respectively, whereas aromatic carbons appeared in the region of 118 ppm to 130 ppm. The  
197 signal at 165 ppm is the characteristic signal for imine carbon (>C=N) (Fig. S3-S6). Mass  
198 spectral analysis of starting azoaldehyde and **H<sub>2</sub>L** exhibit *m/z* peaks at 279.00 and 551.21,  
199 respectively (Fig.S7-S9). Mass spectrum of **H<sub>2</sub>L** in the presence of Al<sup>3+</sup> shows peak *m/z* at 575.25  
200 which is corresponding to [(Al(L))<sup>+</sup> (Fig. S10).

201 The structure of both azoaldehyde and **H<sub>2</sub>L** has been established by single crystal X-ray  
202 diffraction measurement. Crystals of both the organic compound are obtained from slow

203 evaporation of CH<sub>2</sub>Cl<sub>2</sub>-CH<sub>3</sub>OH solvent mixture. The ORTEP plot of azoaldehyde and **H<sub>2</sub>L** are  
204 shown in Fig. 1 and Fig. 2 respectively and the selected bond parameters are listed in Table 2.  
205 The azoaldehyde crystallizes with a monoclinic space group Cc. It has almost planer structure  
206 where the azo nitrogen atoms and the aromatic ring containing -CHO, -OCH<sub>3</sub>, -OH groups  
207 belong in the same plane. The remaining aromatic ring deviates from the plane to a small extent  
208 (10.71°). The N=N and C=O (-CHO) bond distances are 1.252Å and 1.224Å respectively which  
209 are similar with other reported values.<sup>50</sup> Aromatic C-C bond distances observed around 1.368 -  
210 1.404 Å. In solid state it form 2D network through intermolecular hydrogen bonding between the  
211 phenoxy o-atom and aromatic hydrogen atom and edge to edge π-π stacking interaction between  
212 two phenyl rings. The distances are 2.704 Å and 3.70 Å respectively (Fig. S11).

213 The Schiff base ligand (**H<sub>2</sub>L**) crystallizes with a triclinic space group  $P\bar{1}$ . The azo -N=N-  
214 bond distances are 1.217 Å and 1.268 Å respectively. The average C=N- bond distance is 1.287  
215 Å. The C-C bond distances of the phenyl rings found around 1.342-1.472 Å, whereas aliphatic  
216 C-C bond distances appear around 1.511-1.531 Å. The propylenic part of the Schiff base, N3-  
217 C15-C16-C17-N4, is to some extent puckered due to the sp<sup>3</sup> hybridization of the saturated  
218 portion of the ligand. The bond angle (C15-C16-C17, 114.35(6)° deviates appreciably from its  
219 ideal value. OH group of the both of phenyl ring are placed 180° apart from each other to  
220 minimize the dipole-dipole interaction. In the Schiff base ligand each azo nitrogen atoms, imine  
221 atoms and the aromatic ring containing -CHO, -OCH<sub>3</sub>, -OH groups belong in the same plane.  
222 The remaining aromatic ring deviates from the plane significantly by an angle of 83.03°.The  
223 molecule dimerizes in solid state through intermolecular H-bonding between the two -OH  
224 groups as well as between the -OH group and imine nitrogen atom. -O-H...O bond distance is  
225 3.879Å whereas -O-H...N (imine) bond distance is 2.357Å (Fig. S12).

226 Structural parameters are again calculated from optimized structure using DFT where  
227 bond distances are marginally elongated by 0.0023–0.0435 Å and bond angles varies within the  
228 range of 0.05–1.39° (Table 2) compare with the X-ray crystallographic data. Therefore, the  
229 optimized results are helpful to explain the electronic structure and electronic properties of the  
230 ligand **H<sub>2</sub>L**.

### 231 **Absorption study**

232 The UV–Vis spectrum of the Chemosensor **H<sub>2</sub>L** was recorded at 25°C in aqueous buffer-  
233 acetonitrile solution (1:100 v/v, HEPES buffer at pH 7.4) which exhibited well-defined bands at  
234 290 and 388 nm respectively. In order to study the binding property of **H<sub>2</sub>L** toward Al<sup>3+</sup> ion,  
235 UV–Vis spectra of **H<sub>2</sub>L** (10 μM) in the presence of various concentrations of Al<sup>3+</sup> (0–10 μM)  
236 were recorded at room temperature, as shown in Fig. 3. It has been observed that old peaks  
237 disappear and two new absorption bands emerge at 286 and 375 nm respectively, and its  
238 absorbance gradually increases with the gradual addition of Al<sup>3+</sup> and it gets saturated upon  
239 addition of 1.0 equivalents of Al<sup>3+</sup> keeping the concentration of **H<sub>2</sub>L** fixed at 10 μM. These  
240 observations indicate the coordination of **H<sub>2</sub>L** with one eq. of Al<sup>3+</sup> (Fig. 3).

### 241 **Al<sup>3+</sup> ion sensing by fluorescence studies**

242 The fluorescence property of **H<sub>2</sub>L** was also investigated in HEPES buffer solution at pH 7.4 at  
243 room temperature. The ligand, **H<sub>2</sub>L**, emits weakly at 510 nm when excited at 388 nm and the  
244 fluorescence quantum yield is (Φ= **H<sub>2</sub>L**) 0.00939. Upon excitation at 290 nm we observed  
245 similar emission at 510 nm. Low fluorescence intensity of free **H<sub>2</sub>L** probably due to photo  
246 induced electron transfer process (PET) caused by electron delocalization from the two  
247 phenoxido oxygen atoms to the π-conjugated system of two aromatic rings, >C=N– group and  
248 azo (-N=N-) group. Here presence of extended delocalization through azo group (-N=N-)

249 initiates appearance of fluorescence pick at 510 nm. After gradual addition of  $\text{Al}^{3+}$  ion with  
250 various concentrations (0-10  $\mu\text{M}$ ), a significant changes in emission spectra have been noticed.  
251 In presence of metal ion the emission band of  $\text{H}_2\text{L}$  is blue shifted to 478 nm. This result also  
252 confirms a high sensitivity of the receptor towards  $\text{Al}^{3+}$ . A plot of fluorescence intensities at 478  
253 nm ( $I_{478}$ ) vs concentration of aluminum has been given in Fig. 4. : inset. It shows that sensing  
254 character of  $\text{H}_2\text{L}$  ( $I_{478}$ ) increases with the increasing concentration of  $\text{Al}^{3+}$  and a clear bend of the  
255 curve was observed at 1.0 equivalent of added  $\text{Al}^{3+}$  which prove 1:1 stoichiometry of the  
256  $[\text{Al}(\text{L})]^+$ . Such type of sigmoid curve reflects nature of interaction between the organic probe and  
257  $\text{Al}^{3+}$  ion. Upon addition of 1-3  $\mu\text{M}$  of  $\text{Al}^{3+}$  to 10  $\mu\text{M}$  solution of chemosensor, very small change  
258 of fluorescence intensity of the probe at wavelength 478 nm has been observed. Upon gradual  
259 addition of  $\text{Al}^{3+}$  from 4-9  $\mu\text{M}$  fluorescence intensity increases from 300-1400 a.u. which  
260 indicates strong interaction between Schiff base ligand and  $\text{Al}^{3+}$ . Maximum increase of  
261 fluorescence intensity has been observed upto 10  $\mu\text{M}$  addition of  $\text{Al}^{3+}$ . This clearly indicates a  
262 1:1 binding between  $\text{H}_2\text{L}$  and  $\text{Al}^{3+}$ . The curve became a plateau with further addition of  $\text{Al}^{3+}$ ,  
263 when further increment in fluorescence intensity has not been observed.

264 Enhancement in fluorescence intensity of  $\text{H}_2\text{L}$  in presence of  $\text{Al}^{3+}$  probably due to  
265 elimination of photoinduced electron transfer (PET) in free  $\text{H}_2\text{L}$  followed by chelation  
266 enhancement effect (CHEF) through the co-ordination of azomethine-N and phenolic-O to metal  
267 ion (Scheme 2). Hence, upon complexation a large CHEF effect is observed because of the rigid  
268 framework and thereby obstructs the PET process. Thus, upon coordination with  $\text{Al}^{3+}$  ion, the  
269 PET process within the ligand system would become weak by reduction of either the electron-  
270 accepting ability of the  $\pi$ -conjugated system of two aromatic rings,  $>\text{C}=\text{N}-$  and azo ( $-\text{N}=\text{N}-$ )  
271 groups or the electron-donating ability of the phenoxido oxygen atoms.

272 Fluorescence intensity of **H<sub>2</sub>L** has been examined with various cations ( $\text{Cd}^{2+}$ ,  $\text{Co}^{2+}$ ,  $\text{Cr}^{3+}$ ,  
273  $\text{Cu}^{2+}$ ,  $\text{Fe}^{3+}$ ,  $\text{Hg}^{2+}$ ,  $\text{K}^+$ ,  $\text{Mn}^{2+}$ ,  $\text{Na}^+$ ,  $\text{Ni}^{2+}$ ,  $\text{Sn}^{2+}$ ,  $\text{Zn}^{2+}$ ,  $\text{Ag}^+$ ,  $\text{Fe}^{2+}$  and  $\text{Ca}^{2+}$ ) using their nitrate salt. (Fig.  
274 S13). Fig. S13 shows that only  $\text{Al}^{3+}$  can induce significant enhancement of fluorescence  
275 intensity. Interestingly, upon addition of different metal ions the intensity of the fluorescence  
276 either remains unchanged or weakened. In presence of  $\text{Co}^{2+}$ ,  $\text{Cr}^{3+}$ ,  $\text{Cu}^{2+}$ ,  $\text{K}^+$ ,  $\text{Mn}^{2+}$ ,  $\text{Na}^+$ ,  $\text{Ni}^{2+}$ ,  $\text{Fe}^{2+}$   
277 and  $\text{Ca}^{2+}$  metal ions emission intensity remain almost unchanged and in presence of  $\text{Fe}^{3+}$   $\text{Cd}^{2+}$ ,  
278  $\text{Sn}^{2+}$ ,  $\text{Ag}^+$ ,  $\text{Hg}^{2+}$ , and  $\text{Zn}^{2+}$  metal ions emission intensity slightly increased (Fig. S13). A  
279 competition assay of **H<sub>2</sub>L** in the presence of  $\text{Al}^{3+}$  and other metal ions have also been studied and  
280 presented in Fig. 5, which prove that fluorescence enhancement due to  $\text{Al}^{3+}$  nullify the possible  
281 interference of other metal ions. Upon addition of different anions viz.  $\text{S}_2\text{O}_3^{2-}$ ,  $\text{S}^{2-}$ ,  $\text{SO}_3^{2-}$ ,  $\text{SO}_4^{2-}$ ,  
282  $\text{SCN}^-$ ,  $\text{N}_3^-$ ,  $\text{AsO}_4^{3-}$ ,  $\text{PO}_4^{3-}$ ,  $\text{ClO}_4^-$ ,  $\text{AcO}^-$ ,  $\text{Cl}^-$ ,  $\text{NO}_2^-$ ,  $\text{NO}_3^-$  and  $\text{SCN}^-$  in HEPES buffer at pH 7.4 (Fig.  
283 6) chemosensor **H<sub>2</sub>L** showed no significant fluorescence enhancement. Thus, the probe is an  
284 excellent example of fluorescence chemosensor towards  $\text{Al}^{3+}$  in the presence of different metal  
285 ions.

286 To find out the binding ability of our chemosensor **H<sub>2</sub>L** with  $\text{Al}^{3+}$  ions, binding constant  
287 was calculated using Benesi-Hildebrand equation (Equation 1) involving fluorescence titration  
288 curve.

$$289 \frac{F_{\max} - F_0}{F_x - F_0} = 1 + \left( \frac{1}{K[C]^n} \right) \quad (1)$$

290 Where,  $F_{\max}$ ,  $F_0$  and  $F_x$  are fluorescence intensities of **H<sub>2</sub>L** in the presence of  $\text{Al}^{3+}$  at  
291 saturation, free **H<sub>2</sub>L** and any intermediate  $\text{Al}^{3+}$  concentration at  $\lambda_{\max} = 478$  nm.  $K$  is the  
292 dissociation constant of the complex. Concentration of  $\text{Al}^{3+}$  ions is represented by  $C$  and slope  $n$  is  
293 measured with Hill plot. Then Binding constant ( $K_a$ ) of the complex has been determined using

294 the relation,  $K_a = 1/K_d$ . A plot of  $\frac{F_{max}-F_0}{F_x-F_0}$  vs  $\left(\frac{1}{[M]^n}\right)$  provides the apparent binding constant value  
295 as  $3.31 \times 10^3 \text{ M}^{-1}$  (Fig. S14) ( $n=1.0$ ).<sup>14e</sup> The Job's plot suggests that the ligand **H<sub>2</sub>L** forms 1:1  
296 complex with  $\text{Al}^{3+}$  ions (Fig. S15-16). To determine the binding stoichiometry of **H<sub>2</sub>L** and  $\text{Al}^{3+}$   
297 fluorescence intensity is plotted against different mole fractions of  $\text{Al}^{3+}$  while volume of solution  
298 has remained constant (Fig. S16). Maxima in this plot has been obtained at 0.5 mole fraction,  
299 which suggests about 1 : 1 complex formation of **H<sub>2</sub>L** and  $\text{Al}^{3+}$ . The formation of the complex  
300 further confirmed from ESI Mass spectroscopy. Mass spectrum of **H<sub>2</sub>L** in the presence of  $\text{Al}^{3+}$   
301 shows peak  $m/z$  at 575.25 which is corresponding to cationic part of the complex **1**  $[\text{Al}(\text{L})]^+$  (Fig.  
302 S10). The simulated pattern shows well agreement with that of experimental values (Fig. S17).  
303 Experimental findings (absorption spectra and fluorescence spectra of  $\text{Al}^{3+}$  and other cations and  
304 various anions with chemosensor **H<sub>2</sub>L**) clearly revealed that **H<sub>2</sub>L** has strong affinity towards  
305  $\text{Al}^{3+}$ . Here  $\text{Al}^{3+}$  acts as strong Lewis acid which can simply binds with two phenoxido oxygen  
306 atoms and two imine nitrogen atoms of the Schiff base ligand. Thus the chemosensor acts as a  
307 tetradentate N<sub>2</sub>O<sub>2</sub> donor ligand and form a distorted tetrahedral complex with  $\text{Al}^{3+}$ . Comparable  
308 size of the inner-cavity of the chemosensor and  $\text{Al}^{3+}$  i.e. suitable size and high charge density of  
309  $\text{Al}^{3+}$  permits strong interaction between **H<sub>2</sub>L** and the metal ion.

310 Fluorescence decay behavior of **H<sub>2</sub>L** and complex **1** has been studied. Decay curves have  
311 been given in Supplementary Information (Fig. 7). Fluorescence life time of **H<sub>2</sub>L** is found to  
312 quite low (1.324 ns). Fluorescence life time of **H<sub>2</sub>L** in presence of one eq. of  $\text{Al}^{3+}$ , increases up  
313 to 2.058 ns. Quantum yields of **H<sub>2</sub>L** and complex **1** are calculated using equation 2 and found to  
314 be very low. These have been measured to be 0.00939 and 0.2133, respectively (Table S1). It is  
315 to be noted that although quantum yield of the complex is low but it is almost ~22 folds greater  
316 compared to free **H<sub>2</sub>L**.

$$\Phi_x = \Phi_s \left( \frac{F_x}{F_s} \right) \left( \frac{A_s}{A_x} \right) \dots\dots\dots(2)$$

318 Where  $F_x$ ,  $F_s$  are the wavelength integrated emission intensities of the samples and reference  
 319 (here reference is Coumarine);  $A_s$ ,  $A_x$  are the optical densities at their corresponding wavelengths  
 320 of excitation.

321 Free ligand upon excitation gives peak at 510 nm. In presence of  $Al^{3+}$  there is a slight blue shift  
 322  $\sim 32$  nm of intensity and emission peak observed at 478 nm. Upon gradual addition of  $Al^{3+}$  to  
 323  $H_2L$  a steady increase in emission intensity at 478 nm has been observed.

324 Sensitivity of  $H_2L$  towards  $Al^{3+}$  has been checked by determining limit of detection  
 325 (LOD) value. The detection limit of the chemosensor was calculated using  $3\sigma$  method<sup>51</sup> and it is  
 326 found to be  $6.93 \times 10^{-9}$  mol/L.<sup>14e</sup> Low LOD value clearly indicate high sensitivity of  $H_2L$  towards  
 327  $Al^{3+}$  ion. In Table S1 some recently published chemosensors for  $Al^{3+}$  ion has been reported along  
 328 with their LOD values.<sup>14a,14e,14g,14i-14o</sup> The reported chemosensor has some advantages over the  
 329 others while it has some draw backs also. Our probe is synthesized by easy single step Schiff  
 330 base condensation process. LOD value is significantly low but not the lowest among all.

331 We also examined fluorescence intensity of  $H_2L$  in absence and presence of  $Al^{3+}$  ion at  
 332 various pH values. We have maintained the pH range from 3 to 10. The solution concentration of  
 333 both  $H_2L$  and  $Al^{3+}$  is 10  $\mu$ M respectively. It has been observed that the chemosensor in free  
 334 condition exhibit very little fluorescence in the acidic condition ( $p^H = 3$  to  $p^H = 7$ ). Fluorescence  
 335 intensity remains unchanged within this  $p^H$  range. Upon increasing pH from 7 to 10 a slight  
 336 increase in fluorescence intensity has been observed. In presence of  $Al^{3+}$ , in acidic condition ( $p^H$   
 337  $= 3$  to  $p^H = 5$ ) fluorescence intensity of the chemosensor remain unchanged due to protonation of  
 338 phenolic hydroxyl group of the ligand, therefore nullifying chelation ability with  $Al^{3+}$ .  $Al^{3+}$  ion



339 sensing abilities were exhibited by the ligand when pH was increased from 5.0 to 8.0. Thus, **H<sub>2</sub>L**  
340 exhibited good fluorescence sensing ability towards Al<sup>3+</sup> ion over a wide range of p<sup>H</sup> (Fig. 8).  
341 Based on the experimental results we can conclude that our chemosensor can be utilized as a  
342 selective fluorescent probe to recognize and distinguish Al<sup>3+</sup> ion in presence of other metal ions  
343 in biological system under physiological condition.

#### 344 **DFT study**

345 Geometry optimization of both **H<sub>2</sub>L** and [Al(L)]<sup>+</sup> has been performed using DFT/B3LYP  
346 method. Mass spectral data analysis has confirmed that the composition of the complex ion is  
347 [Al(L)]<sup>+</sup>. The energy minimized structure of both **H<sub>2</sub>L** and [Al(L)]<sup>+</sup> are shown in Fig 9. Some  
348 selected bond distances and bond angles of the optimized geometries of [Al(L)]<sup>+</sup> are listed in  
349 Table 3. DFT optimized structure reveals that Al<sup>3+</sup> is tetra-coordinated and distorted tetrahedral  
350 in geometry (Fig. 9) where the chemosensor **H<sub>2</sub>L** act as a N2O2 donor center and binds to Al<sup>3+</sup>  
351 through phenolic-O atom and imine-N. Theoretically calculated Al–N(imine) and Al–  
352 O(phenolato) distances are around 1.906Å and 1.326Å, respectively and have been comparable  
353 with similar structure.<sup>14e</sup> It is interesting to mention that upon complex formation the C(15)–  
354 N(3), C(17)–N(4), C(1)–O(1) and C(20)–O(2) bond distances have been significantly elongated  
355 compared to free ligand (Table 3). Theoretical calculations show that electron density in both  
356 HOMO and LUMO of **H<sub>2</sub>L** is mainly distributed on the one part of the aromatic moiety. In metal  
357 complex ion i.e. [Al(L)]<sup>+</sup> electron density in both HOMO and LUMO is allocated over the entire  
358 ligand system. Energy of some selected M.O. of both **H<sub>2</sub>L** and [Al(L)]<sup>+</sup> are given in Table S3.  
359 Mulliken charge distribution shows positive charge on aluminium ion i.e. 1.227343. Mulliken  
360 charge distribution of [Al(L)]<sup>+</sup> is provided in Table S4.

361

362 **TDDFT study**

363 For better understanding of electronic transition, TDDFT calculations were performed  
364 using B3LYP/CPCM method using same basis sets in acetonitrile. The calculated electronic  
365 transitions are given in Table 4. **H<sub>2</sub>L** shows intense absorption band for ligand based n- $\pi$  and  $\pi$ -  
366  $\pi^*$  transitions around 290 nm and 388 nm respectively. The band at 290 nm is due to the  
367 contribution of HOMO-4 $\rightarrow$ LUMO+1 and HOMO $\rightarrow$ LUMO+2 transitions, whereas the band at  
368 388 nm is due to the contribution of HOMO-1 $\rightarrow$ LUMO, HOMO-1 $\rightarrow$ LUMO+1 and  
369 HOMO $\rightarrow$ LUMO transitions. For the L-Al species the intense absorption band around 375 nm  
370 corresponding to HOMO $\rightarrow$ LUMO+1, HOMO $\rightarrow$ LUMO+2, HOMO $\rightarrow$ LUMO+3 and HOMO-2  
371  $\rightarrow$ LUMO transitions (Fig. 10). The spectra calculated for the ligand and L-Al species were  
372 found to be compatible with those obtained experimentally. It is important to mention that in  
373 case of [Al(L)]<sup>+</sup> species all transitions are ligand based.

374 In order to have better understanding of the emission spectrum we have optimized the  
375 triplet state (T1) of both ligand (**H<sub>2</sub>L**) and ([Al(L)]<sup>+</sup>) using same basis set. The emission  
376 wavelength obtained from the computation is very much comparable with that of experimental  
377 data. The data i.e. emission wavelength, emission energies and the nature of the transition  
378 obtained from the computation are given in Table 5. The emission band of the chemosensor  
379 (**H<sub>2</sub>L**) at 510 nm was theoretically obtained at 509.84 nm with major key transitions of  
380 HOMO( $\alpha$ ) $\rightarrow$ LUMO( $\alpha$ )+1(58%), HOMO( $\alpha$ ) $\rightarrow$ LUMO( $\alpha$ )+2(10%) and HOMO( $\beta$ )-1 $\rightarrow$ LUMO( $\beta$ )  
381 (11%) respectively. Whereas for the [Al(L)]<sup>+</sup> the emission band at 478 nm was theoretically  
382 obtained at 476.93 with major key transitions viz. HOMO( $\beta$ ) $\rightarrow$ LUMO( $\beta$ )+2(22%) and  
383 HOMO( $\alpha$ )-1 $\rightarrow$ LUMO( $\alpha$ ) (66%) respectively.

384

385 **NMR studies**

386 Firstly, the chemosensor was well characterized in  $\text{CDCl}_3$  solvent. In  $\text{CDCl}_3$ , imine  
387 proton appeared at 8.43 ppm. Aromatic, aliphatic and methoxy protons appeared at 7.42-7.90  
388 ppm, 3.82-3.84 ppm, 2.25 ppm and 4.01 ppm respectively. Then, we have performed  $^1\text{H}$  NMR  
389 studies of both free  $\text{H}_2\text{L}$  and  $\text{H}_2\text{L}+\text{Al}^{3+}$  in 1:1 molar ratio in  $\text{DMSO-d}_6$  solvent (Fig. 11). The  
390 free chemosensor ( $\text{H}_2\text{L}$ ) exhibit clear peaks for different protons. Imine proton appears at 8.66  
391 ppm. All aromatic protons appear as multiplet at around 7.74-7.30 ppm. Aliphatic protons  
392 appear as multiplet at 3.74 and 2.06 ppm respectively.  $-\text{OCH}_3$  protons appear around 3.74 ppm.  
393 In NMR spectrum signal of aliphatic one  $-\text{CH}_2$  got merged with that the signal of  $-\text{OCH}_3$ .  
394 Unfortunately we could not identify signal for phenolic OH proton due to extensive hydrogen  
395 bonding with solvent molecules. Upon addition of  $\text{Al}^{3+}$  to the chemosensor peak positions of  
396 different protons have changed significantly. Imine protons have shifted notably from 8.66 ppm  
397 to 9.24 ppm indicating coordination of imine nitrogen with aluminium ion. In case of aromatic  
398 and aliphatic protons broadening and overlapping of signals observed. Therefore, formation of  
399 complex **1** in the presence of  $\text{Al}^{3+}$  is strongly evidenced from the NMR spectral studies.

400

401 **Molecular logic gates**

402 The property of this chemosensor encourages us to construct a molecular logic gate with  
403 two binary inputs. The two inputs are  $\text{Al}^{3+}$  and  $\text{Na}_2\text{EDTA}$  where the output is monitored as  
404 change in fluorescence emission spectrum at 478 nm. In absence of both of the inputs there is no  
405 significant change in emission band which implies that the gate is 'OFF'. On addition of  
406  $\text{Na}_2\text{EDTA}$  to the chemosensor there is no significant changes, rather the emission band remained

407 unchanged. So, in presence of Na<sub>2</sub>EDTA the output was considered to be zero. On the other  
408 hand, upon addition of Al<sup>3+</sup> ion solution alone to the chemosensor shows significant  
409 enhancement of the fluorescence intensity at 478 nm and now the output signal is 1 i.e. the gate  
410 is 'ON'. However in presence of both of the inputs the fluorescence intensity is significantly  
411 quenched and the output is zero implying that the gate is 'OFF' again. These studies suggest that  
412 this molecular gate is acting as INHIBIT (INH) logic gate where the inputs and output are Al<sup>3+</sup>,  
413 Na<sub>2</sub>EDTA and change in fluorescence intensity at 478 nm, respectively. It is basically an 'AND'  
414 gate which has one of its inputs is negated. The input which is negated, acts to inhibit the gate.  
415 We can say that the gate will behave like an 'AND' gate only when the negated input is set at a  
416 logic level '0'. Here the negated input is Na<sub>2</sub>EDTA. The respective truth table and pictorial  
417 diagram of the INH logic gate is depicted in Fig.12 and Table 6.

## 418 Conclusion

419 In summary, an azo based low cost, simple, easy to prepare, chemosensor **H<sub>2</sub>L** has been  
420 successfully prepared which is capable of recognizing Al<sup>3+</sup> ion in presence of large number of  
421 other metal ions in HEPES buffer solution (1:100 v/v, HEPES buffer at pH 7.4) at 25 °C. The  
422 chemosensor has been structurally characterized. Fluorescence intensity of the probe has  
423 enhanced by ~61 fold in presence of Al<sup>3+</sup>. The low detection limit of **H<sub>2</sub>L** for Al<sup>3+</sup> (6.93 nM)  
424 suggests that the chemosensor could be a good choice for efficient monitoring of the Al<sup>3+</sup> in real  
425 samples. **H<sub>2</sub>L** forms 1:1 complex with Al<sup>3+</sup> which has been established by <sup>1</sup>H NMR, MS studies  
426 and further supported by DFT calculations. **H<sub>2</sub>L** exhibited good fluorescence sensing ability  
427 towards Al<sup>3+</sup> ion over a wide range of p<sup>H</sup>, therefore, **H<sub>2</sub>L** could be successfully applied to living  
428 cells for detecting Al<sup>3+</sup>. We also establish Molecular logic gates (INH) based on two inputs (Al<sup>3+</sup>

429 and EDTA) and one Output. Thus, this probe could be considered as a potential candidate for  
430 sensing  $Al^{3+}$  in less organic solvents.

431

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439

### 440 **Appendix A. Supplementary data**

441 CCDC 1478332-1478333 contain the supplementary crystallographic data for azoaldehyde and  
442 **H<sub>2</sub>L**. This data can be obtained free of charge via  
443 <http://www.ccdc.cam.ac.uk/conts/retrieving.html>, or from the Cambridge Crystallographic Data  
444 Centre, 12 Union Road, Cambridge CB2 1EZ, UK; fax: (+44) 1223-336-033; or  
445 email: [deposit@ccdc.cam.ac.uk](mailto:deposit@ccdc.cam.ac.uk).

### 446 **Notes and References**

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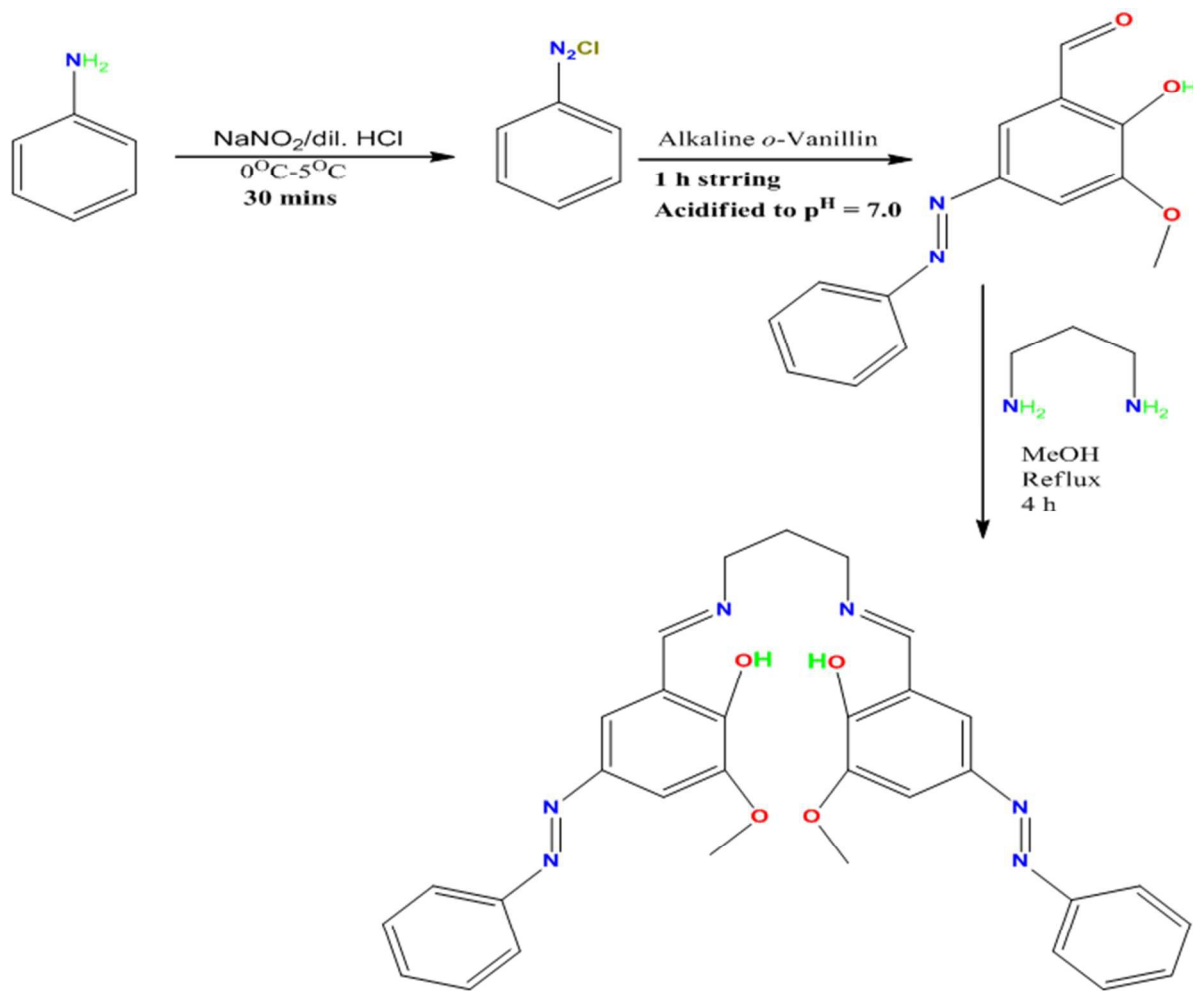


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603 Scheme 1. The route to the syntheses of **H<sub>2</sub>L**.

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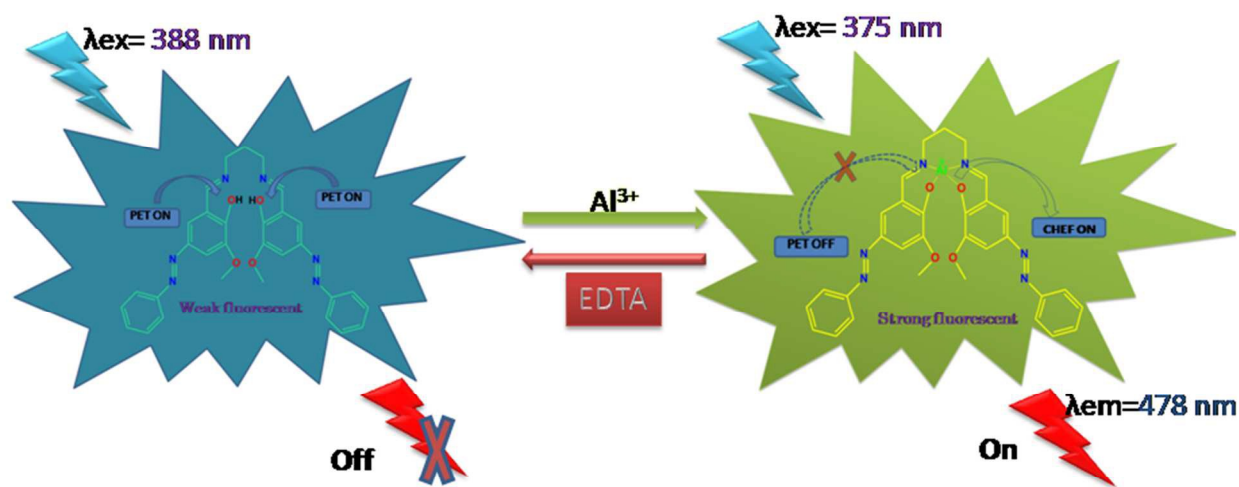
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621 Scheme 2. Schematic diagram for photoinduced electron transfer (PET) in free  $\text{H}_2\text{L}$  and  
 622 chelation enhancement effect (CHEF) in  $[\text{Al}(\text{L})]^+$ .

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634 Table 1. Crystal parameters and selected refinement details for **H<sub>2</sub>L**.

Compound	Azoaldehyde	H <sub>2</sub> L
Empirical formula	C <sub>14</sub> H <sub>12</sub> N <sub>2</sub> O <sub>3</sub>	C <sub>31</sub> H <sub>28</sub> N <sub>6</sub> O <sub>4</sub>
Formula weight	256.26	548.59
Temperature (K)	155(2)	150(2)
Crystal system	Monoclinic	Triclinic
Space group	CC	
<i>a</i> (Å)	10.7683(4)	8.648(2)
<i>b</i> (Å)	13.4401(5)	11.051(3)
<i>c</i> (Å)	8.4892(3)	13.816(4)
$\alpha$ (°)	90.00	91.833(9)
$\beta$ (°)	93.755(2)	97.710(9)
$\gamma$ (°)	90.00	92.902(8)
Volume (Å <sup>3</sup> )	1225.98(8)	1305.8(6)
<i>Z</i>	4	2
<i>D</i> <sub>calc</sub> (g cm <sup>-3</sup> )	1.388	1.395

Absorption coefficient ( $\text{mm}^{-1}$ )	0.100	0.095
$F(000)$	536	576
$\theta$ Range for data collection ( $^\circ$ )	2.43-26.40	2.326-23.559
Reflections collected	4910	12084
Independent reflections / $R_{\text{int}}$	2204/0.0313	3806/ 0.0726
Observed reflections [ $I > 2\sigma(I)$ ]	1980	2308
Data / restraints / parameters	2204/2/175	3806/0/373
Goodness-of-fit on $F^2$	1.029	1.752
Final indices [ $I > 2\sigma(I)$ ]	R1 = 0.0331 wR2 = 0.0812	R1 = 0.1611 wR2 = 0.4280
$R$ indices (all data)	R1 = 0.0378 wR2 = 0.0842	R1 = 0.2089 wR2 = 0.4619
Largest diff. peak / hole ( $\text{e } \text{\AA}^{-3}$ )	0.150/-0.120	0.968/-0.620

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639 Table 2. Selected bond lengths (Å) and bond angles (°) for **Azoaldehyde** and **H<sub>2</sub>L**

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Azoaldehyde			H <sub>2</sub> L		
	X-ray	Calculated		X-ray	Calculated
C3-O2	1.340	1.3450	N1-N2	1.22(1)	1.2645
C8-O1	1.224	1.2264	N5-N6	1.267(9)	1.2647
N1-N2	1.252	1.2611	C15-N3	1.29(1)	1.2853
C2-O3	1.362	1.3631	C17-N4	1.28(1)	1.2845
			C15-C16-C17	114.4(7)	115.79
			C1-O1	1.259	1.275
			C2-O2	1.286	1.297

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657 Table 3. Selected bond lengths (Å) and bond angles (°) of DFT optimized structure of [Al(L)]<sup>+</sup>.  
658 (B3LYP/6-31+G(d) basis set)

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	Calculated		Calculated		Calculated
N1-N2	1.26019	C1-O1	1.32767	Al-O1	1.7584
N5-N6	1.26320	C2-O2	1.32668	Al-O2	1.7553
C15-N3	1.31355	Al-N3	1.90639		
C17-N4	1.31661	Al-N4	1.92372		
C15-C16-C17	115.79	C15-C16-C17	116.50435		

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670 Table 4. Electronic transition calculated by TDDFT using B3LYP/CPCM method in Acetonitrile  
671 solvent of Ligand ( $\text{H}_2\text{L}$ ) and  $[\text{Al}(\text{L})]^+$ .

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	$E_{\text{excitation}}$ (ev)	$\lambda_{\text{excitation}}$ (nm)	Osc. Strength (f)	Key transition	Character
Ligand ( $\text{H}_2\text{L}$ )	3.60	344.43	0.1608	HOMO-4→LUMO+1(42%)	$\pi(\text{L}) \rightarrow \pi^*(\text{L})$
	3.57	346.63	0.0469	HOMO→LUMO+2 (29%)	$\pi(\text{L}) \rightarrow \pi^*(\text{L})$
	3.19	388.28	0.902	HOMO-1→LUMO (56%)	$\pi(\text{L}) \rightarrow \pi^*(\text{L})$
	3.14	394.53	0.6101	HOMO-1→LUMO+1(48%)	$\pi(\text{L}) \rightarrow \pi^*(\text{L})$
	3.019	410.64	0.0149	HOMO→LUMO (71%)	$\pi(\text{L}) \rightarrow \pi^*(\text{L})$
$[\text{Al}(\text{L})]^+$	3.17	390.79	0.2576	HOMO-1→LUMO+1(86%)	$\pi(\text{L}) \rightarrow \pi^*(\text{L})$
	3.15	393.75	0.0254	HOMO→LUMO+2 (82%)	$\pi(\text{L}) \rightarrow \pi^*(\text{L})$
	3.02	411.01	0.0011	HOMO-1→LUMO (86%)	$\pi(\text{L}) \rightarrow \pi^*(\text{L})$
	2.95	419.85	0.6153	HOMO→LUMO (89%)	$\pi(\text{L}) \rightarrow \pi^*(\text{L})$

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676 Table 5. Emission spectrum calculated by TDDFT using B3LYP basis set for Ligand (**H<sub>2</sub>L**) and  
677 [**Al(L)**]<sup>+</sup>.

	E <sub>excitation</sub> (ev)	λ <sub>emission</sub> (nm)	Excited State	Osc. Strength (f)	Key transition
Ligand <b>(H<sub>2</sub>L)</b>	2.43	509.84	5	0.0116	HOMO(α)→LUMO(α)+1(58%) HOMO(β)-1→LUMO(β) (11%) HOMO(α)→LUMO(α)+2 (10%)
[ <b>Al(L)</b> ] <sup>+</sup>	2.17	476.93	8	0.2476	HOMO(α)-1→LUMO(α)(66%) HOMO(β)→LUMO(β) +2 (22%)

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691 Table 6. Truth Table

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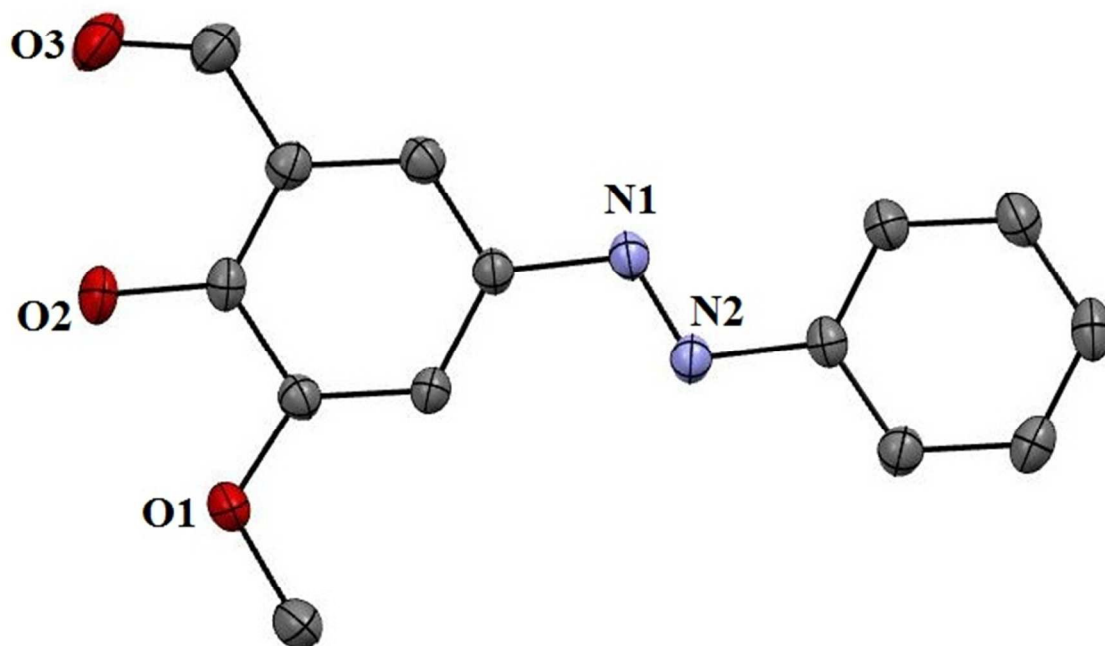
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IN1 (Al <sup>3+</sup> )	IN2 (EDTA)	Output (Emission at 478 nm)
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1	0	1
1	1	0



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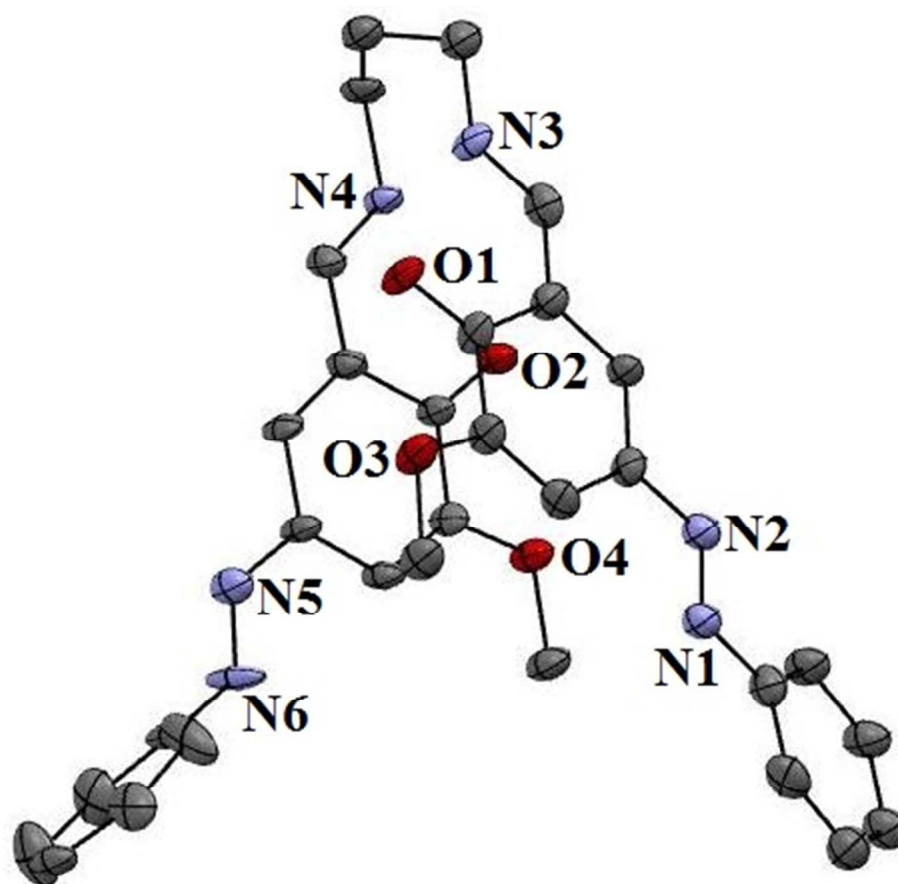
724 Fig. 1. Ortep view of azoaldehyde. Atoms are shown as 30% thermal ellipsoids. H atoms are  
725 omitted for clarity.

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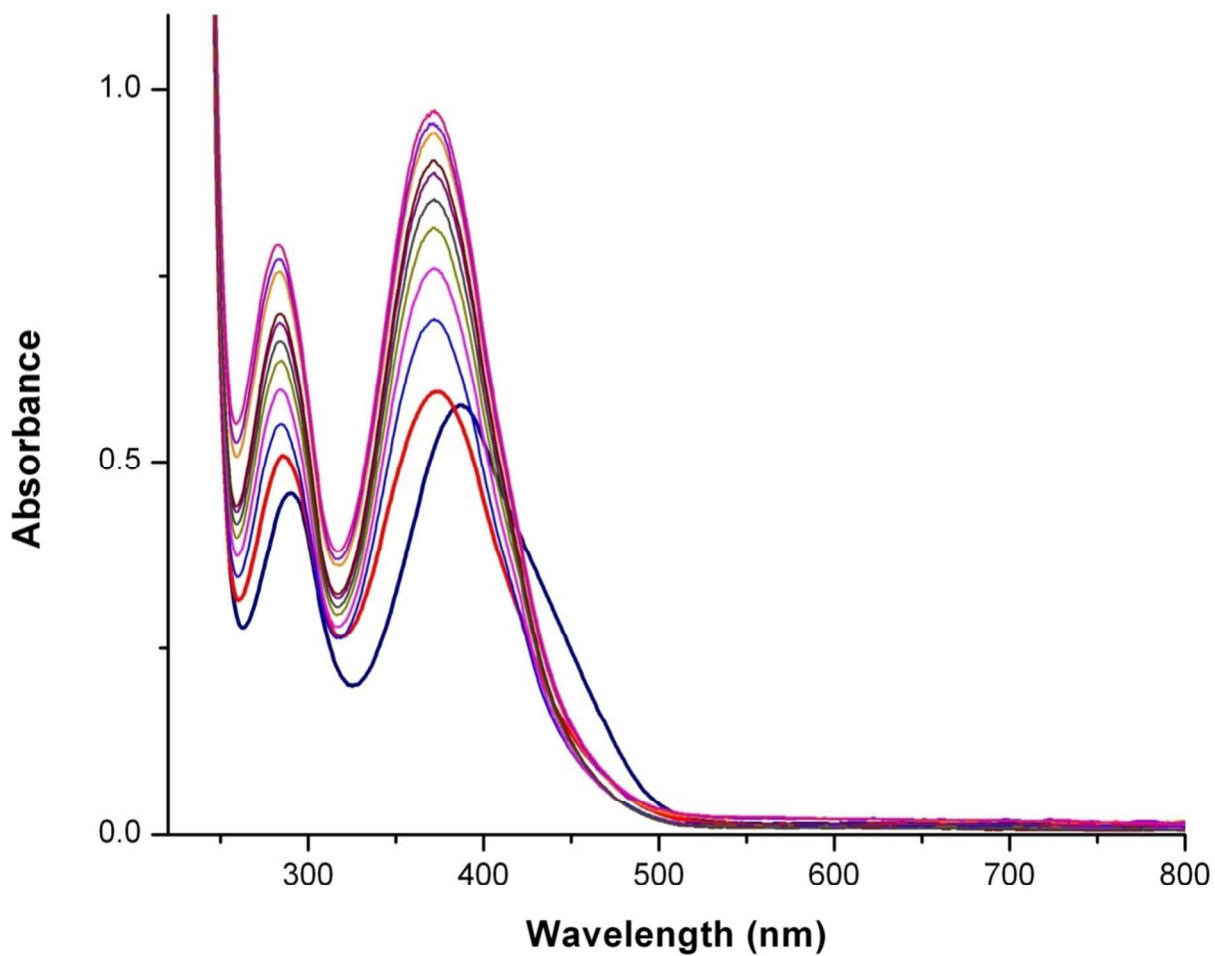
Fig. 2. Ortep view of the ligand ( $H_2L$ ). Atoms are shown as 30% thermal ellipsoids. H atoms are omitted for clarity.

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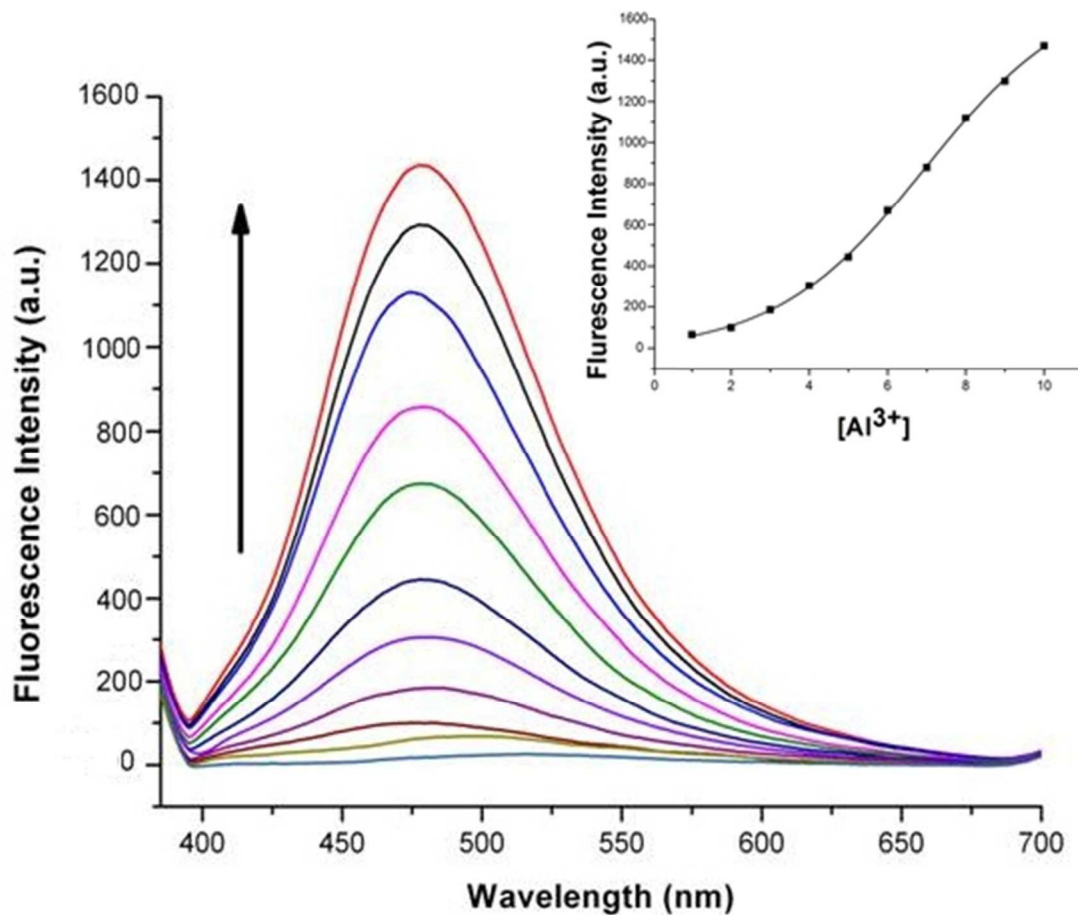
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761 Fig. 3. Absorption titration of **H<sub>2</sub>L** (10 μM) with gradual addition of Al<sup>3+</sup>, 0-10 μM in MeCN/762 HEPES buffer at p<sup>H</sup> 7.4.

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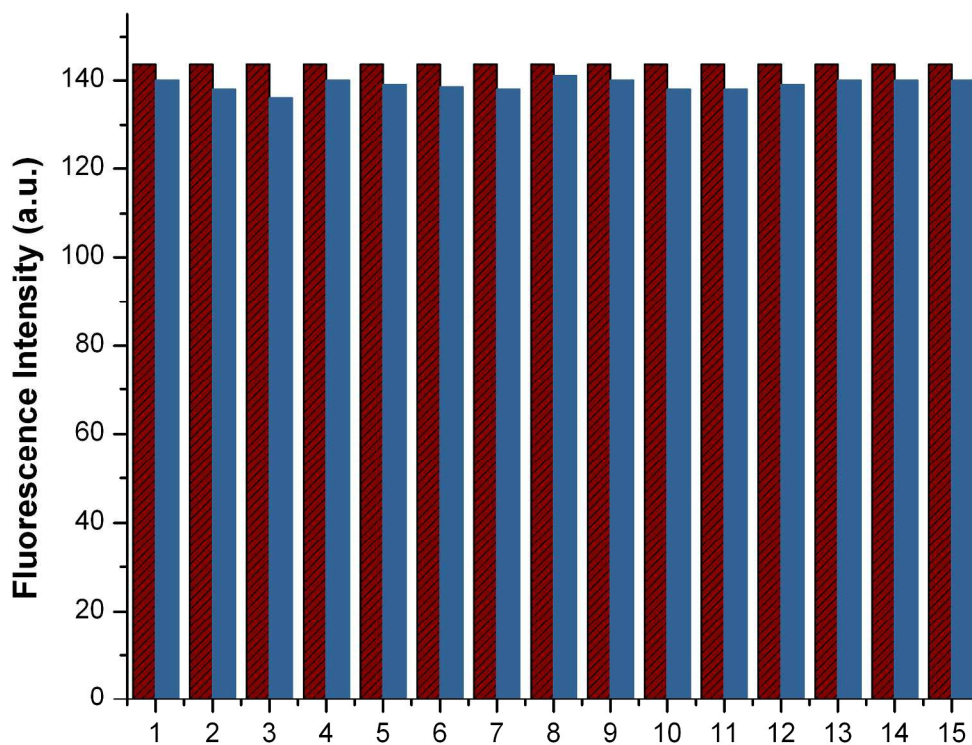
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Fig. 4. Fluorescence titration of  $\text{H}_2\text{L}$  ( $10 \mu\text{M}$ ) in HEPES buffer at  $\text{pH} = 7.4$  by gradual addition of  $\text{Al}^{3+}$  ( $0$ – $10 \mu\text{M}$ ) with  $\lambda_{\text{em}} = 478 \text{ nm}$  ( $5/5$  slit). Inset: non-linear plot of fluorescence intensity vs. concentration of  $\text{Al}^{3+}$  ion.



802 Fig. 5. Relative fluorescence intensity profile of  $[H_2L-Al^{3+}]$  system in the presence of different  
 803 cations in HEPES buffer at pH 7.4.  $H_2L(10 \mu M) + Al^{3+}(10 \mu M) + M^{n+}(500 \mu M)$ , where  $M^{n+}=(1-$   
 804  $Cd^{2+}, 2-Co^{2+}, 3-Cr^{3+}, 4-Cu^{2+}, 5-Fe^{3+}, 6-Hg^{2+}, 7-K^+, 8-Mn^{2+}, 9-Na^+, 10-Ni^{2+}, 11-Sn^{2+}, 12-Zn^{2+}, 13-$   
 805  $Ag^+, 14-Fe^{2+}, 15-Ca^{2+})$ .

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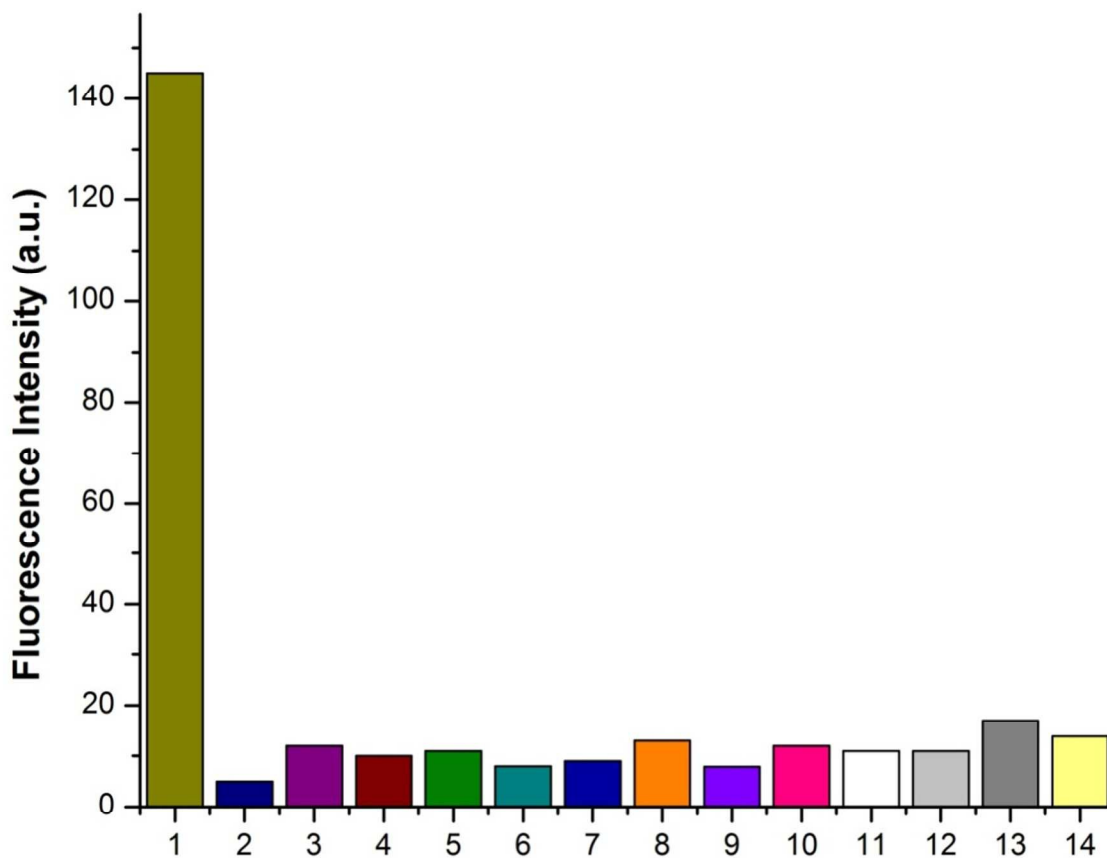
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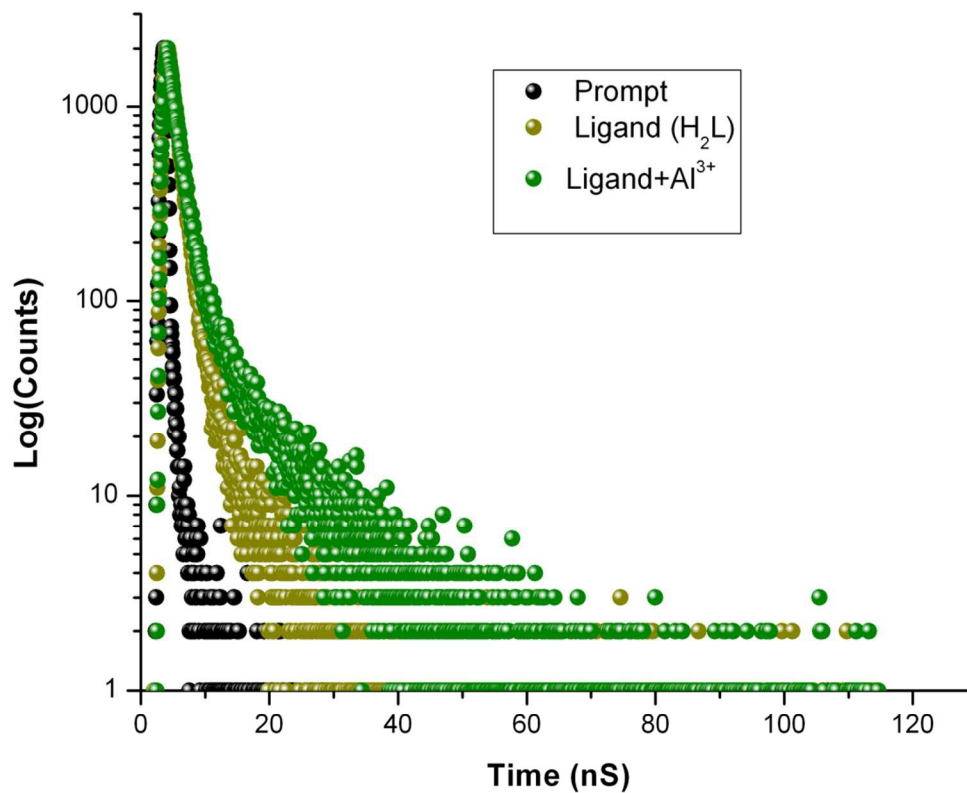
822 Fig. 6. Relative fluorescence intensity profile of  $\text{H}_2\text{L}$  (10  $\mu\text{M}$ ) in the presence of various  
823 common anions (50  $\mu\text{M}$ ) in HEPES buffer at pH 7.4.

824 1-  $\text{H}_2\text{L} + \text{Al}^{3+}$ , 2 – 14  $\text{H}_2\text{L} + \text{Anions}$ , Anions = 2- $\text{S}_2\text{O}_3^{2-}$ , 3- $\text{S}^{2-}$ , 4- $\text{SO}_3^{2-}$ , 5-  $\text{SO}_4^{2-}$ , 6- $\text{SCN}^-$ , 7- $\text{N}_3^-$ ,  
825 8- $\text{AsO}_4^{3-}$ , 9- $\text{PO}_4^{3-}$ , 10- $\text{ClO}_4^-$ , 11- $\text{OAc}^-$ , 12- $\text{Cl}^-$ , 13- $\text{NO}_2^-$ , 14- $\text{NO}_3^-$

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844 Fig. 7. Time-resolved fluorescence decay curves (logarithm of normalized intensity vs time in

845 ns) of H<sub>2</sub>L in the absence (●) and presence (●) of Al<sup>3+</sup> ion, (●) indicates decay curve for846 the scattered ( $\lambda_{ex}$ = 478 nm).

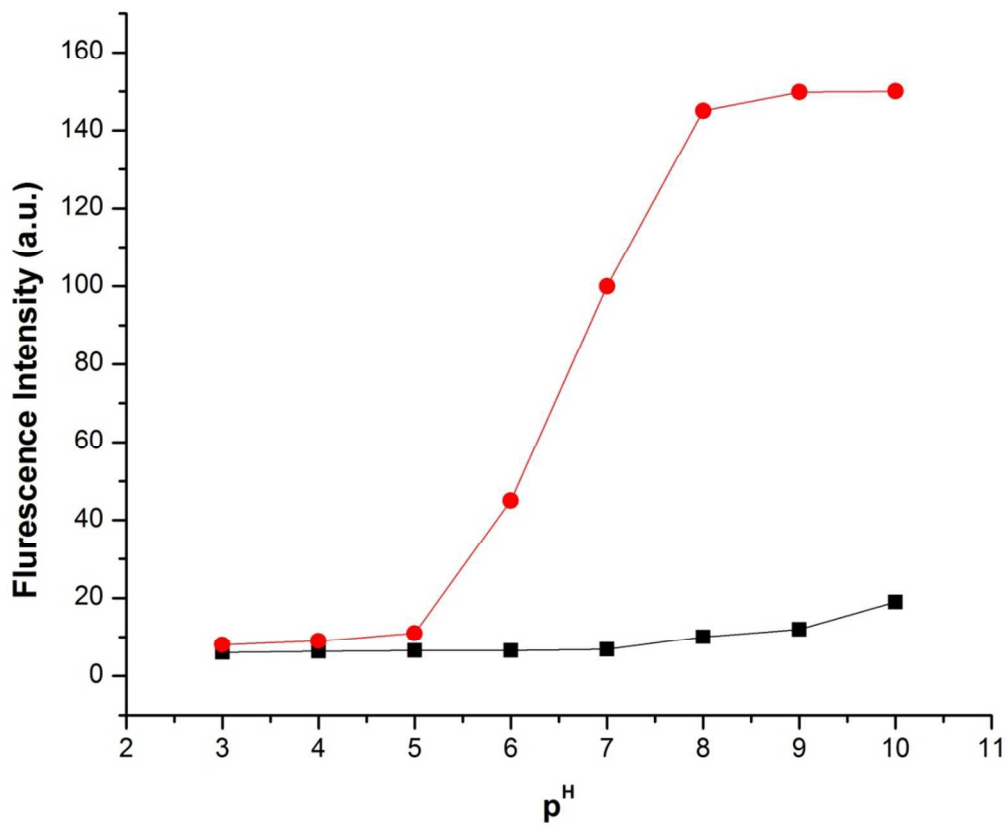
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867 Fig. 8. Fluorescence intensity of H<sub>2</sub>L (black box; 10 μM) in the absence and presence of Al<sup>3+</sup> ion

868 (red circle, 10 μM) ion at various pH values in HEPES buffer.

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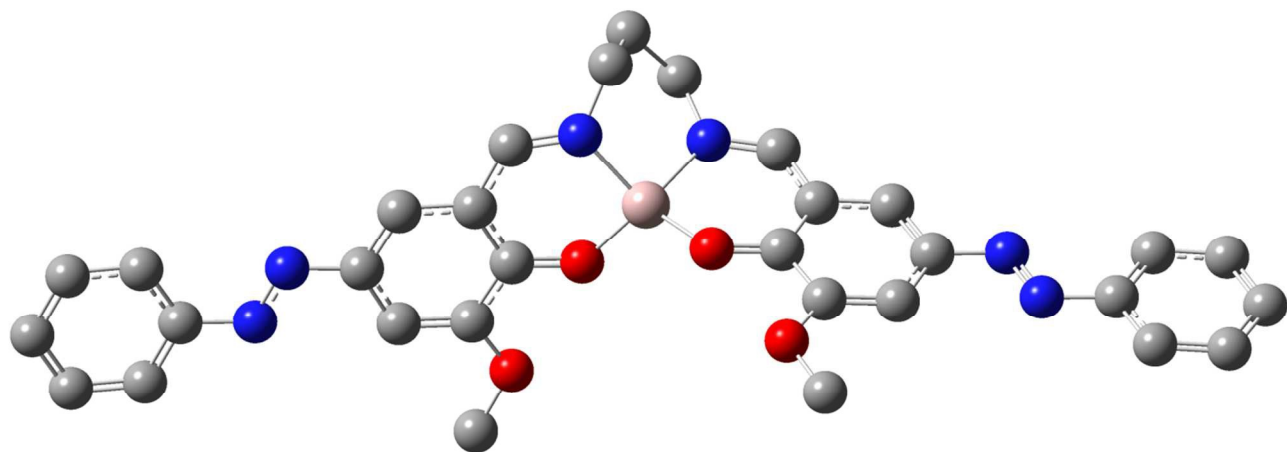
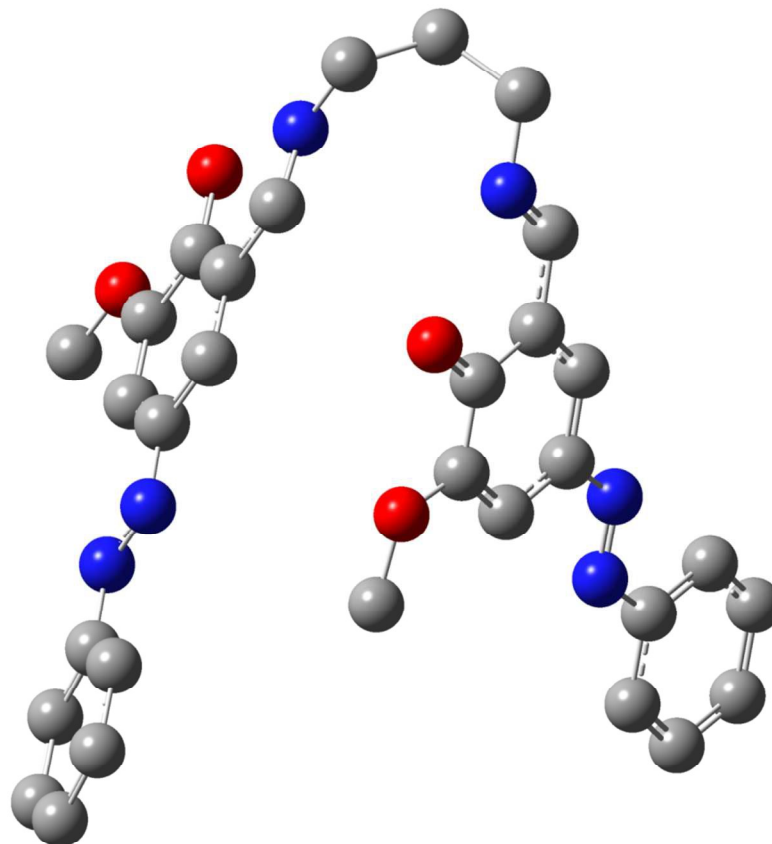
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890 Fig.9. DFT optimized structures of **H<sub>2</sub>L** and **[Al(L)]<sup>+</sup>**. H-atoms are omitted for clarity.

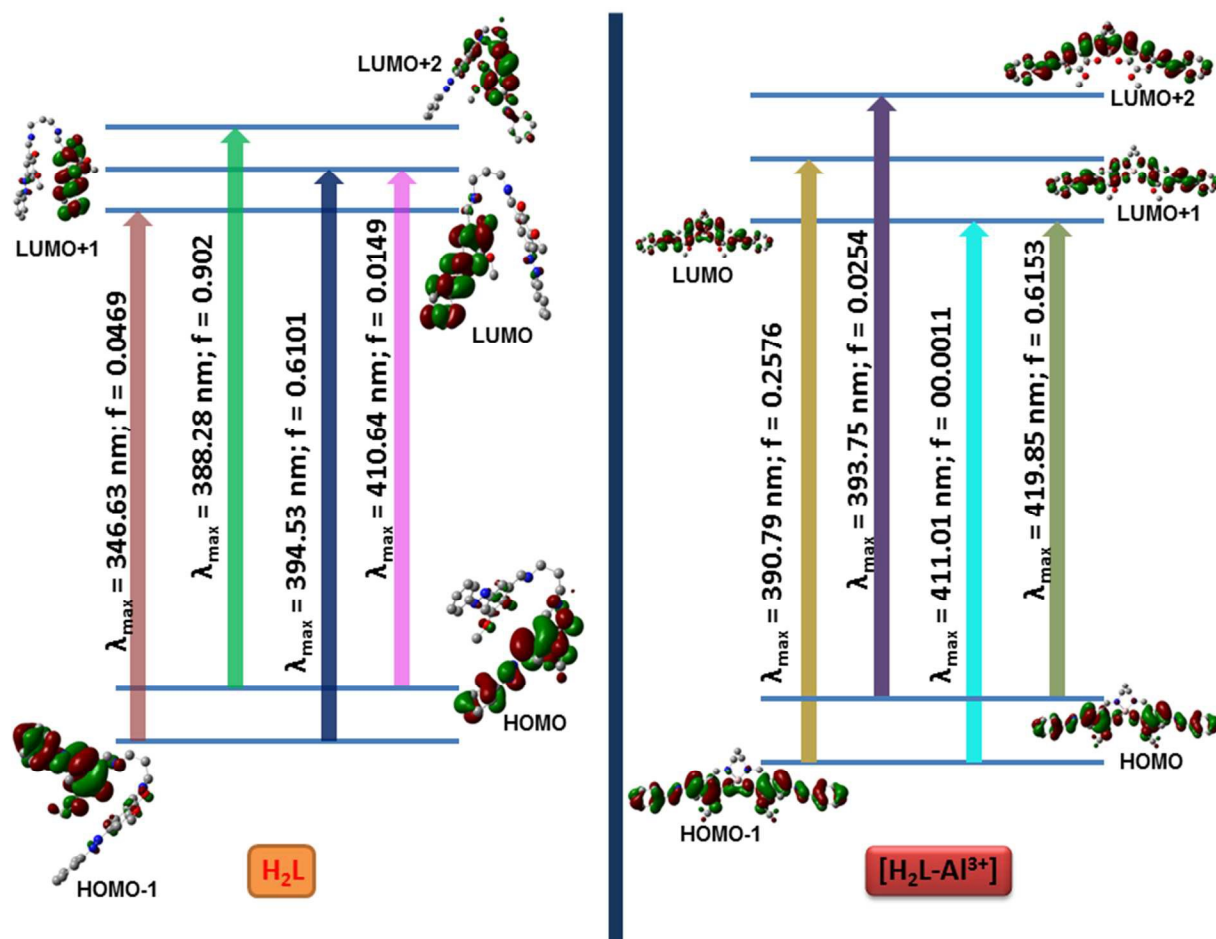
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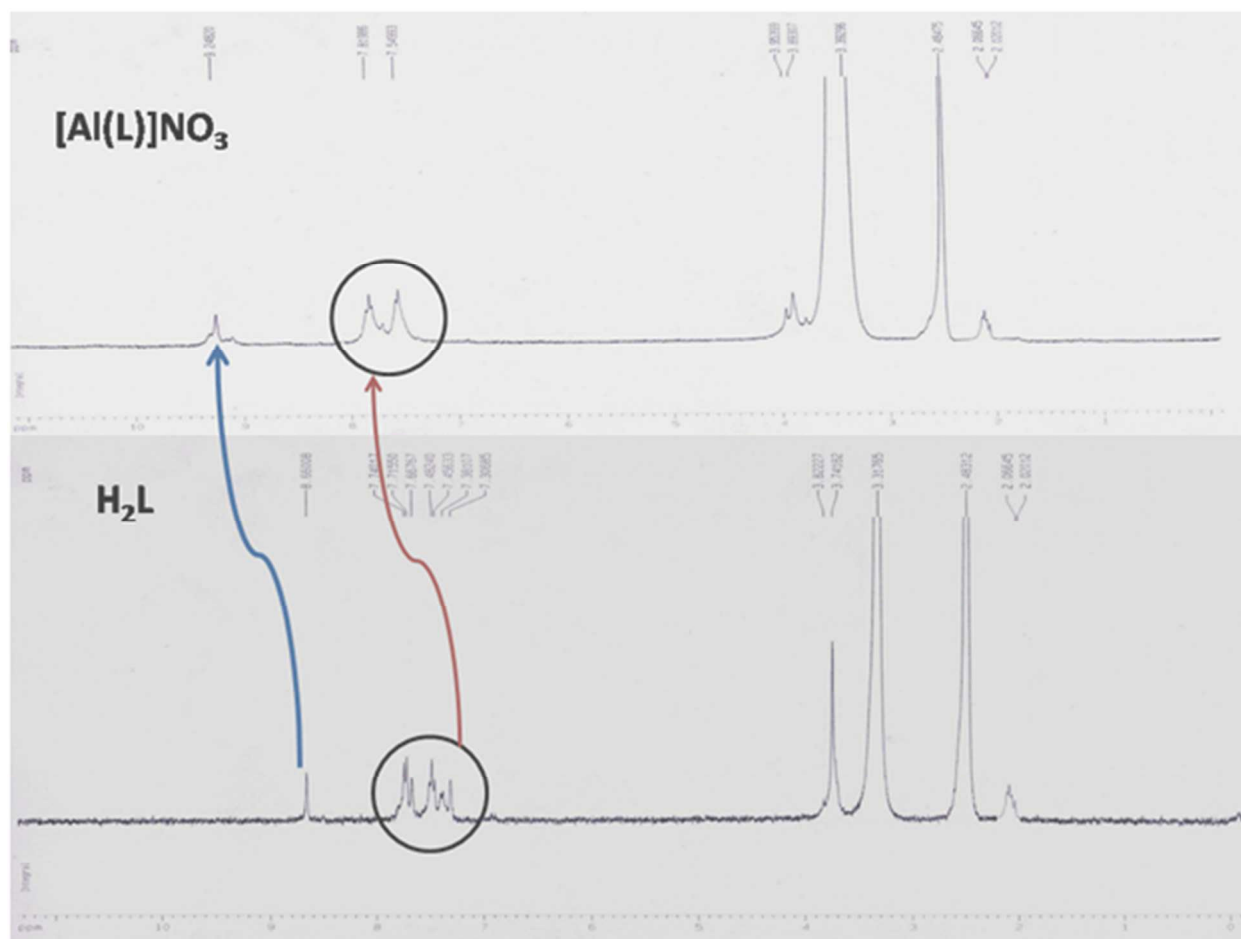
Fig.10. Pictorial representation of key transitions of  $\text{H}_2\text{L}$  and  $[\text{Al}(\text{L})]^+$ .

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909 Fig. 11. NMR titration of  $H_2L$  in presence of 1 eq. of  $Al^{3+}$  ion.

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917 Fig. 12. Pictorial representation of Logic Gate.

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# A Robust Fluorescent Chemo sensor for Aluminium Ion Detection Based on Schiff base Ligand with Azo Arm and Application in Molecular Logic Gate

Saikat Banerjee<sup>a</sup>, Paula Brandão<sup>b</sup> and Amrita Saha<sup>a,\*</sup>

Herewith, we have reported a new Schiff base chemosensor with azo arm for Al<sup>3+</sup> ions. The sensor has been well characterized using different techniques like single crystal x-ray, NMR, IR, UV etc. It shows excellent selectivity over other cations and anions. Detection limit of the sensor was found to be 6.93 nM.

## Graphical Abstract (Pictogram)

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Saikat Banerjee<sup>a</sup>, Paula Brandão<sup>b</sup> and Amrita Saha<sup>\*a</sup>

