

MP0701 COST Workshop, *Electrospun Nanofibres Composite Materials*
Antalya, Turkey, February 21-22, 2012

- *Invited oral communication* -

Electrospun nanofibrous mats outstanding materials for biotechnology applications

J.A. Lopes da Silva

Department of Chemistry, University of Aveiro, Portugal

jals@ua.pt

Electrospun nanofibrous mats

The great advantages

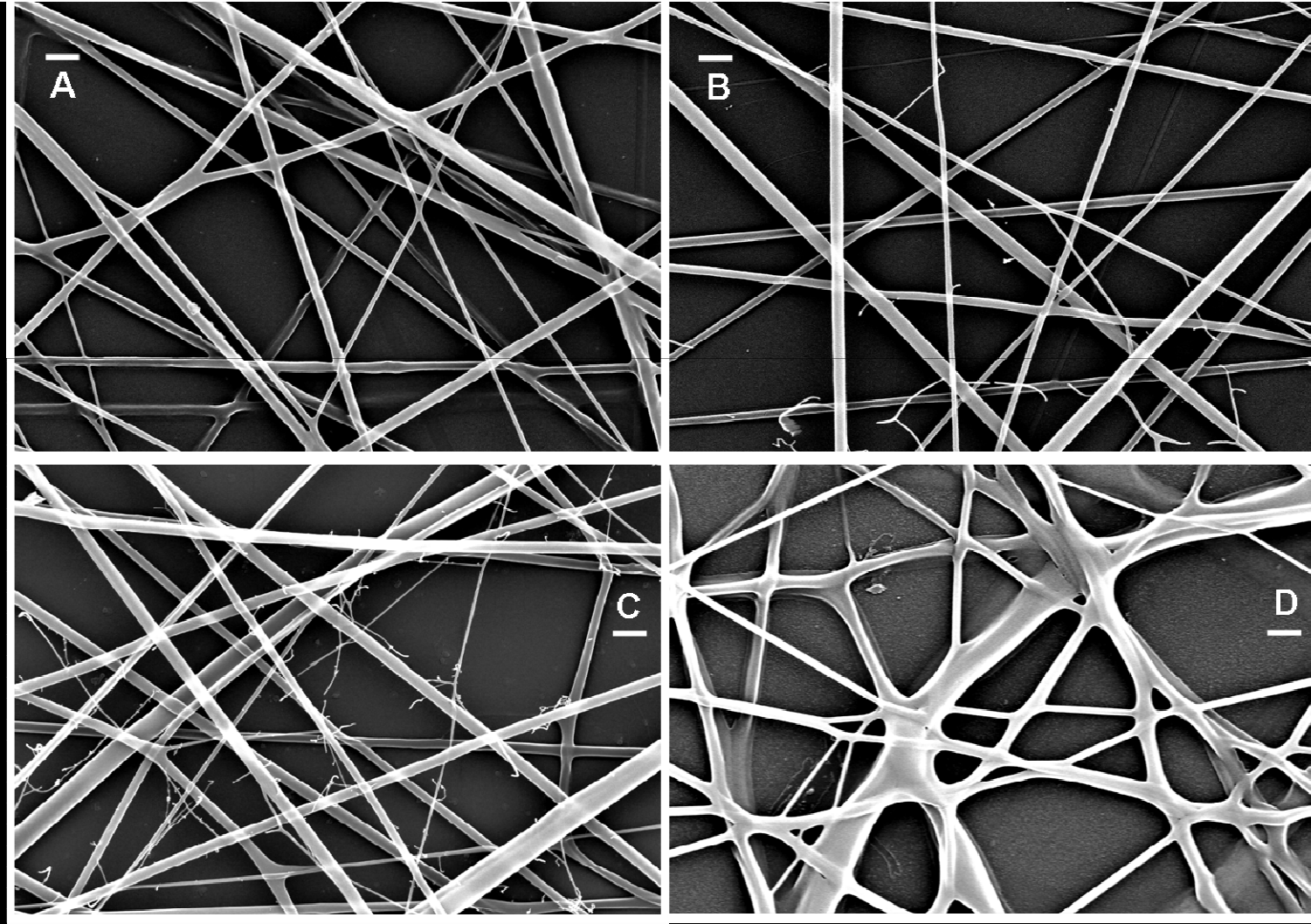
- Nano- and micro-sized fibers
- From random to aligned fiber arrays
- High porosity and interconnected pores
- Prone to manipulation
- Bulk and surface composition versatility

'Our' materials

- Focus on cheap and/or renewable materials, such as polyethylene terephthalate (PET), microbial polyesters (PHBV), polyvinyl alcohol (PVA), chitosan, pectins, cellulose ...

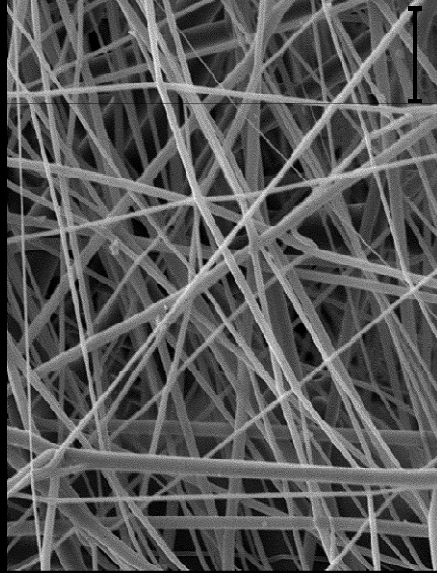
Veirinho, B., Ribeiro-do-Valle, R.M., Lopes da Silva, J.A. (2011)
Materials Letters, 65, pp. 2216–2219.

Electrospun poly (3-hydroxybutyrate-co-hydroxyvalerate) / chitosan blend fibers

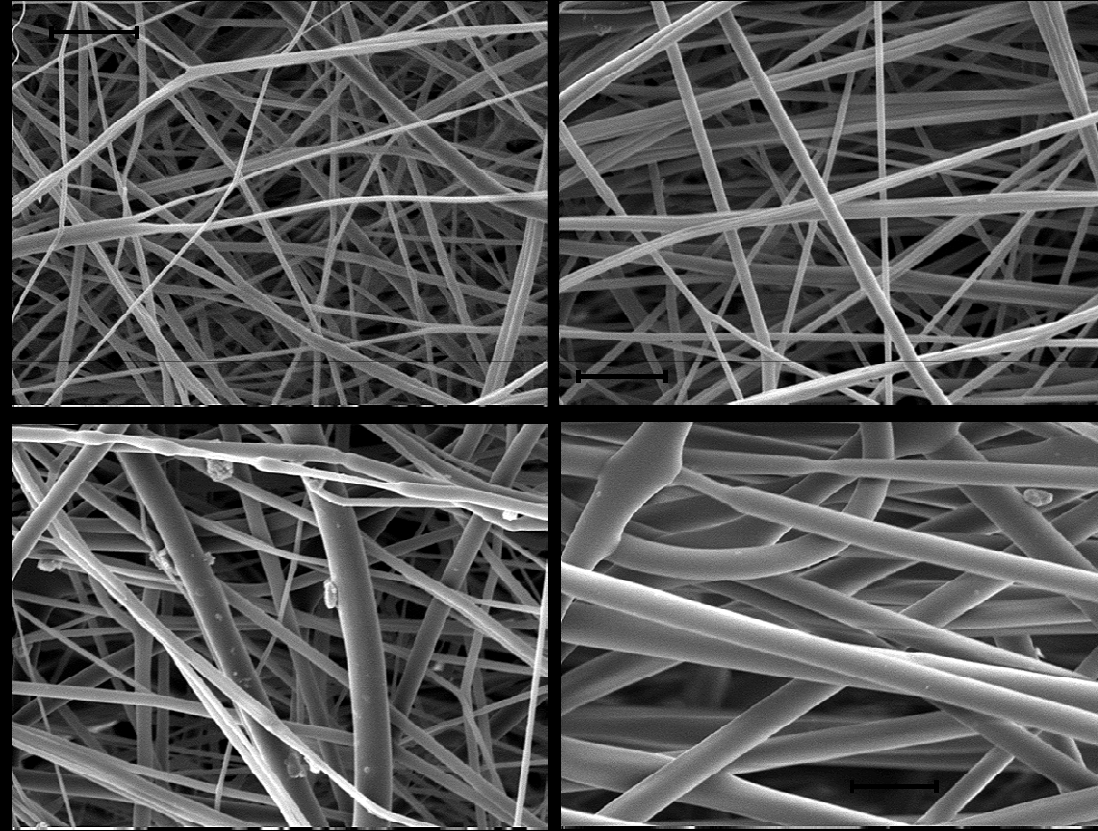


Electrospun PET and PET/chitosan blend fibers

Lopes da Silva, J.A., Veleirinho, B., Delgado I. (2009)
Journal of Nanoscience and Nanotechnology, 9, pp. 3798-3804.

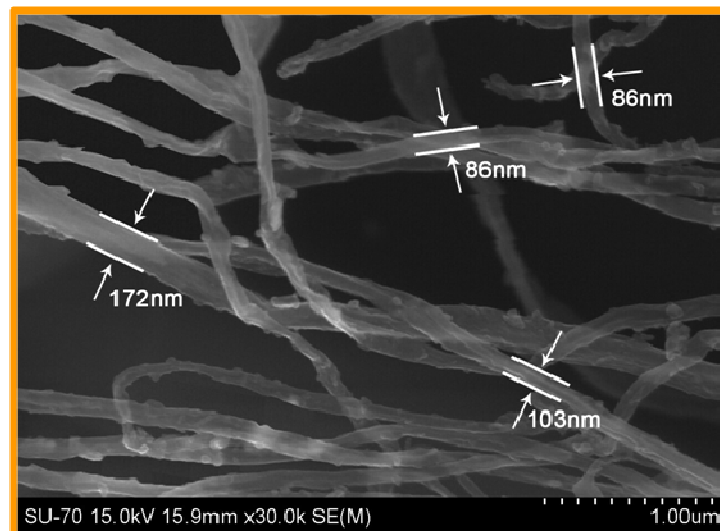
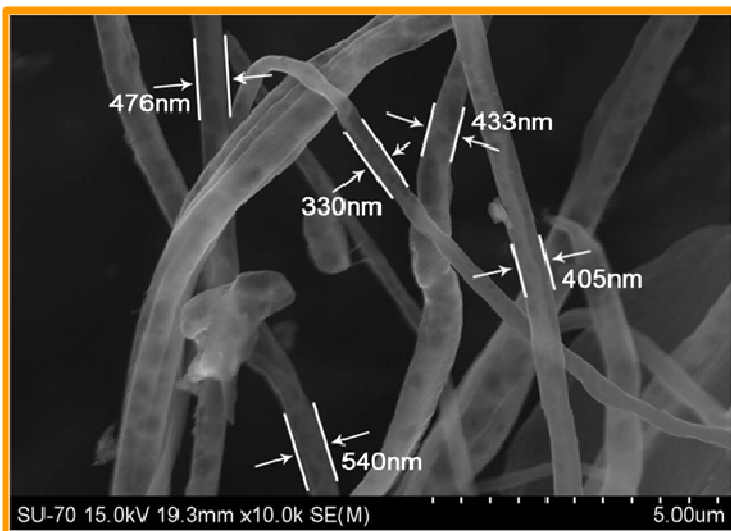
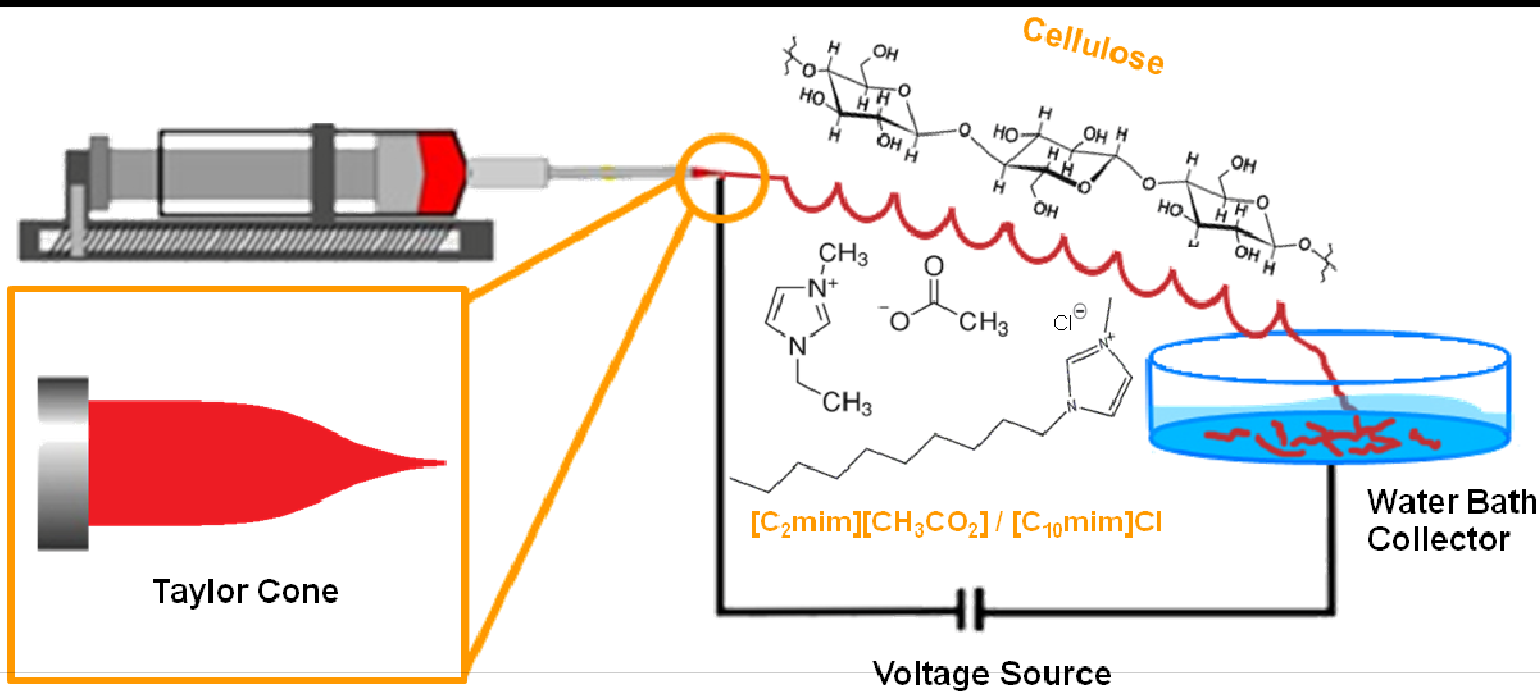


PET



PET/chitosan blend fibers
(different chitosan concentration and molecular weight)

Freire *et al.*, Electrospun nanosized cellulose fibers using ionic liquids at room temperature. *Green Chem.*, 2011, 13, 3173.



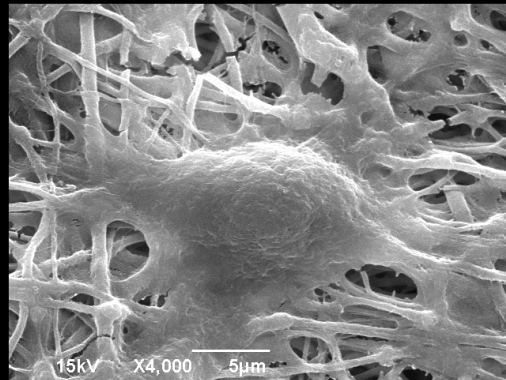
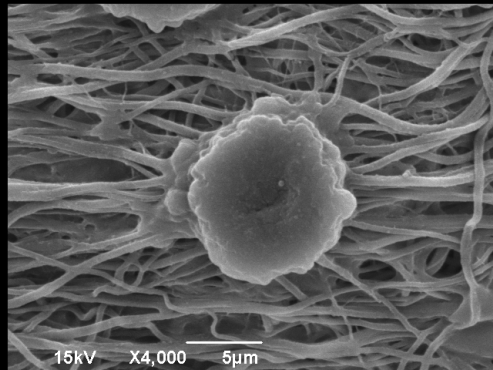
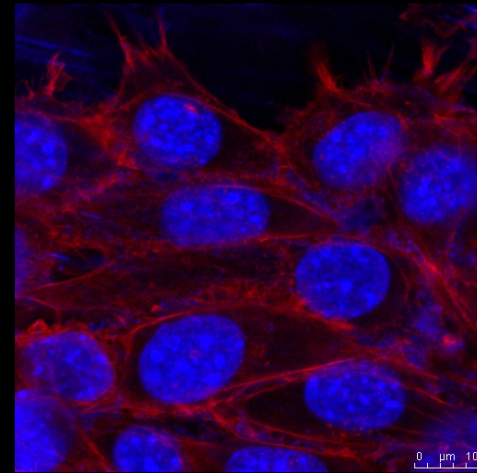
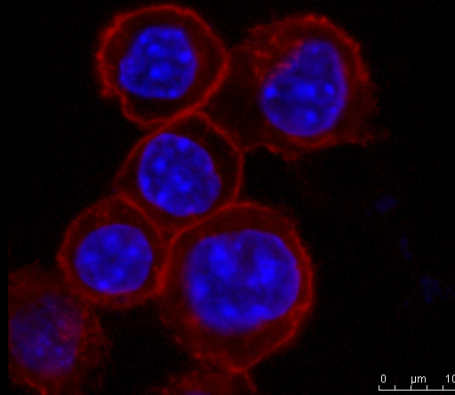
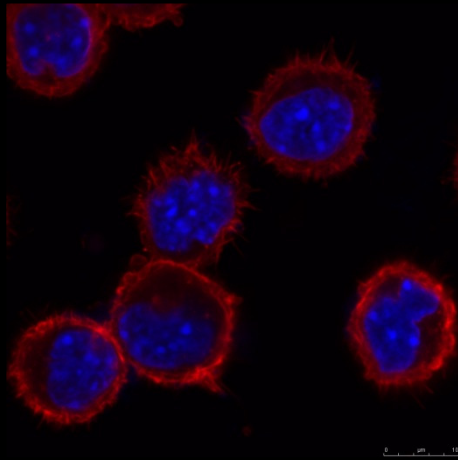
Electrospun nanofibrous mats

'Our' applications

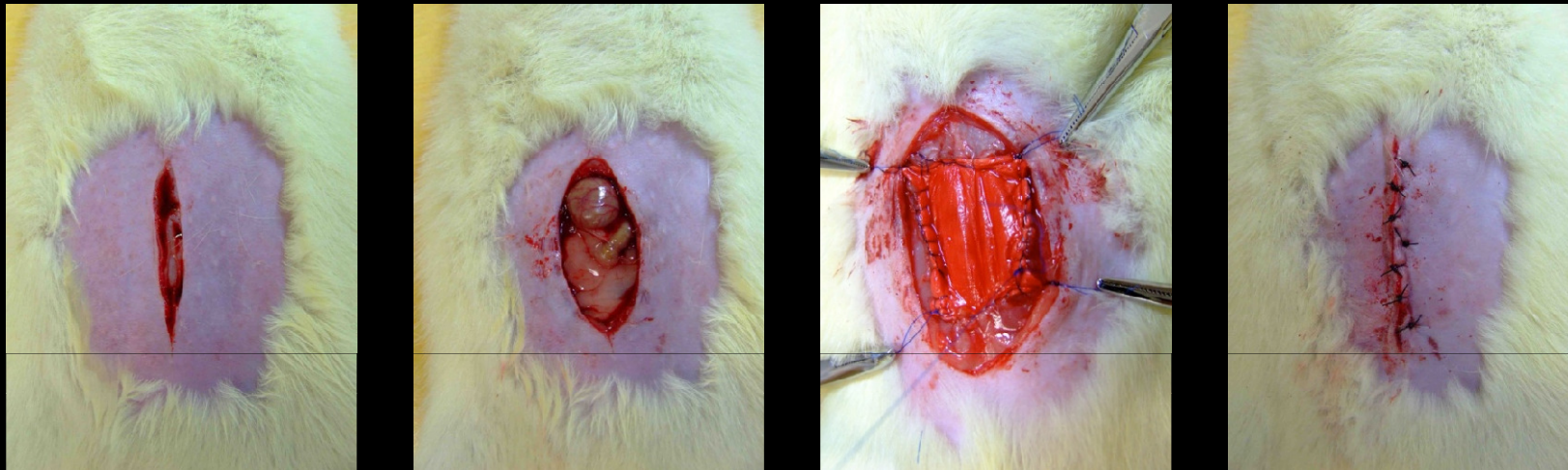
- **Cell-scaffold interactions**
Potential membranes for wound dressing
Potential for hernia meshes
- **Separation/filtration applications**
Potential membranes for juice clarification
- **Fibrous mats as supports for bioactive compounds immobilization/encapsulation**
Potential supports for enzyme immobilization

Electrospun fibrous mats as wound dressing membranes

- *In vitro* growth of fibroblasts on PHBV/chitosan scaffolds



Polymer meshes for hernia repair



In vivo tests using inlay implantation in rodent animal models

Materials

Commercial meshes: Marlex© (polypropylene), Dracon© (polyethylene terephthalate)

PET mesh – *average fiber diameter* = 340 μm

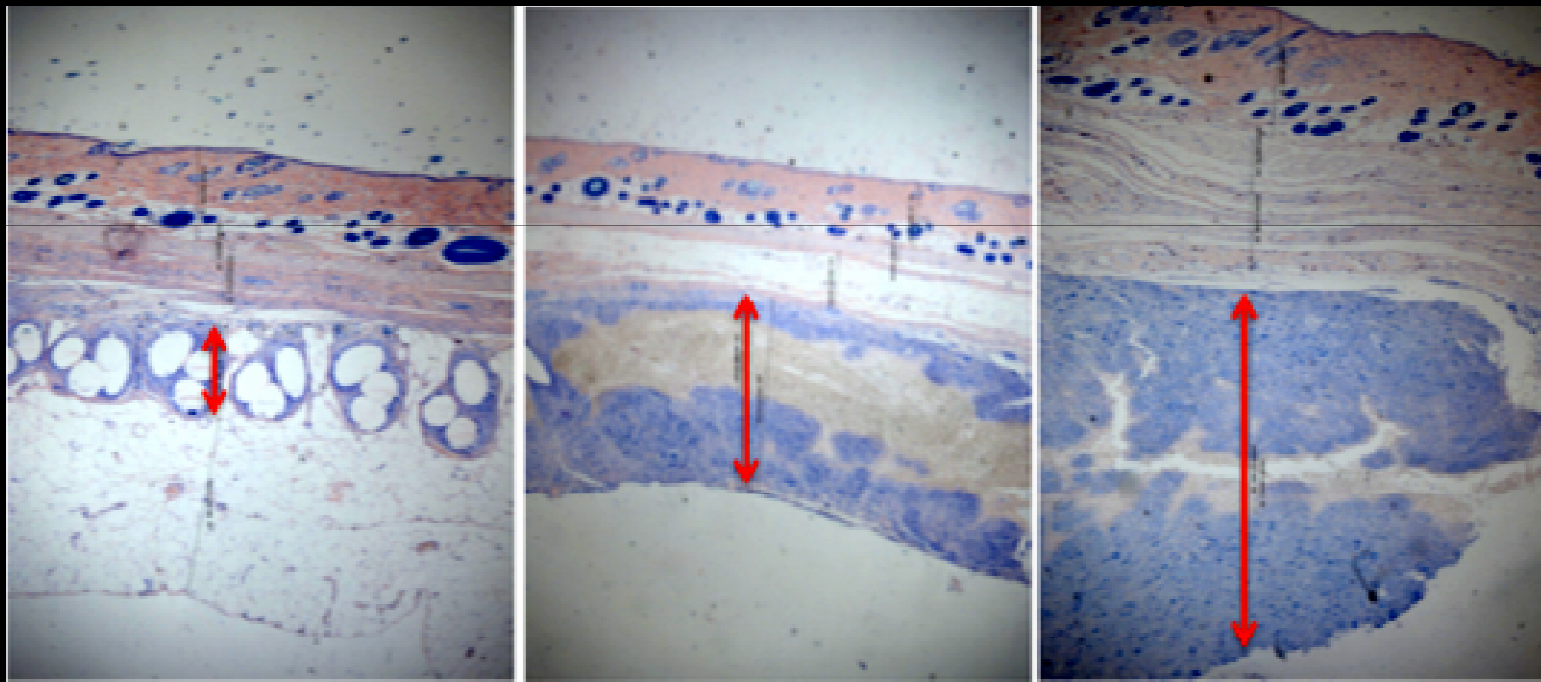
PP mesh – *average fiber diameter* = 208 μm

Electrospun PET mat – *average fiber diameter* = 0.7 μm

Electrospun PET+chitosan mat – *average fiber diameter* = 1.2 μm

Histological evaluation Day 30

Changes on foreign body granuloma



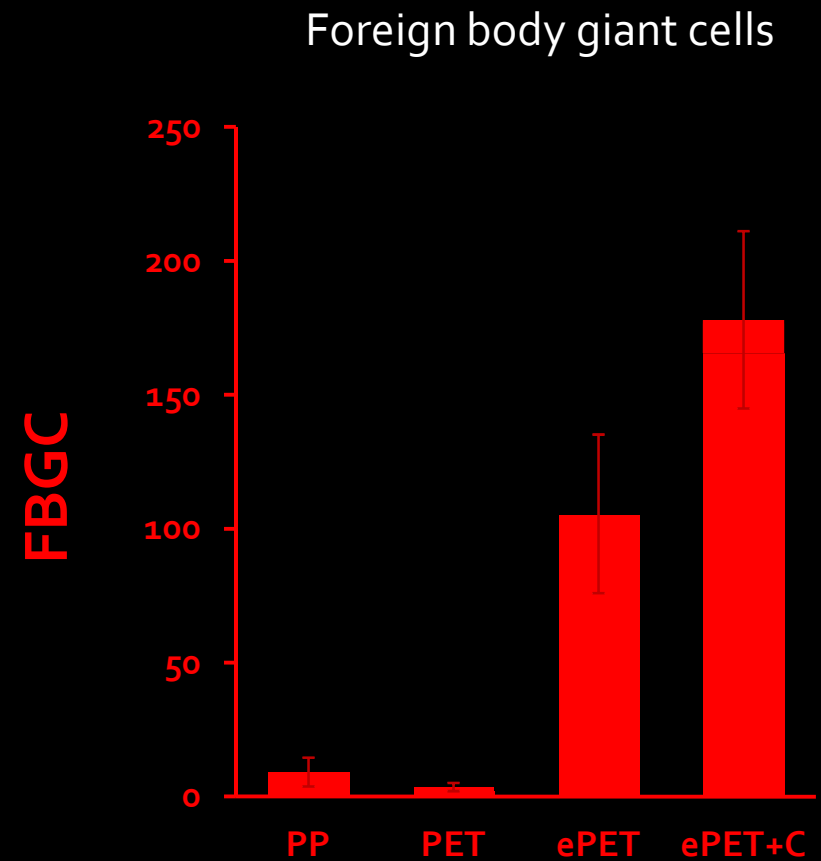
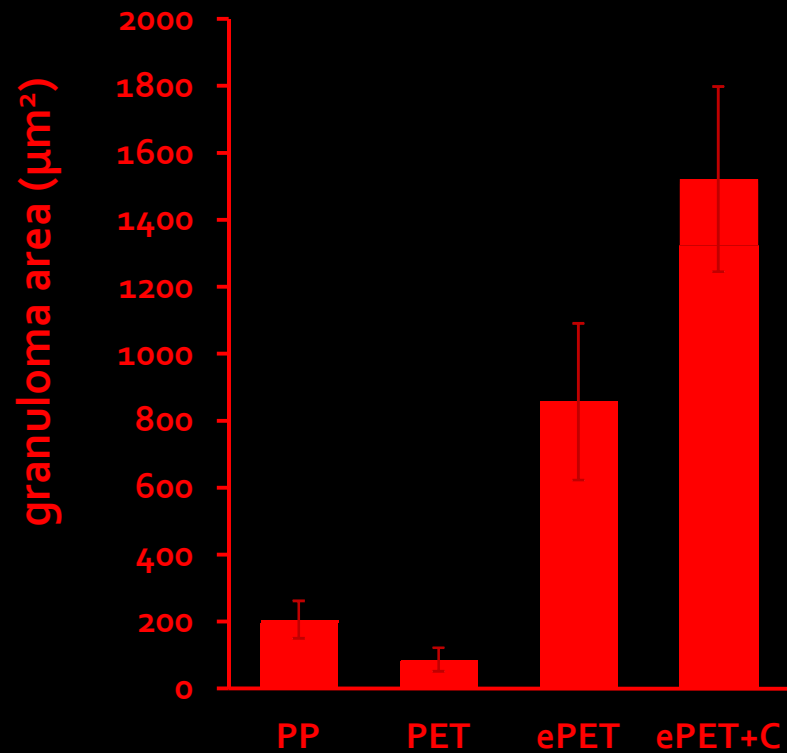
PP (Marlex©)

electrospun PET

electrospun
PET+Chitosan

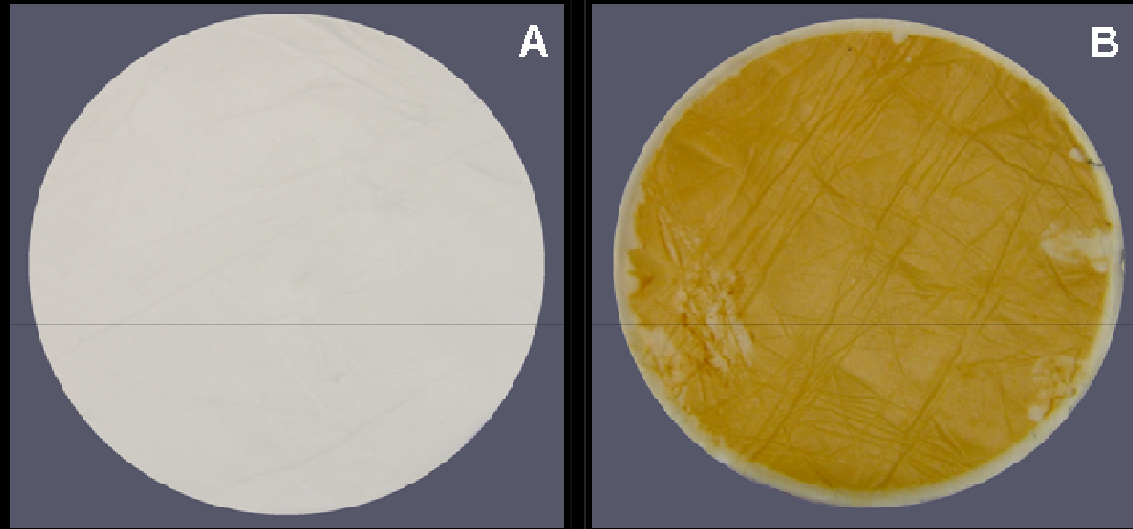
Histological evaluation

Day 30



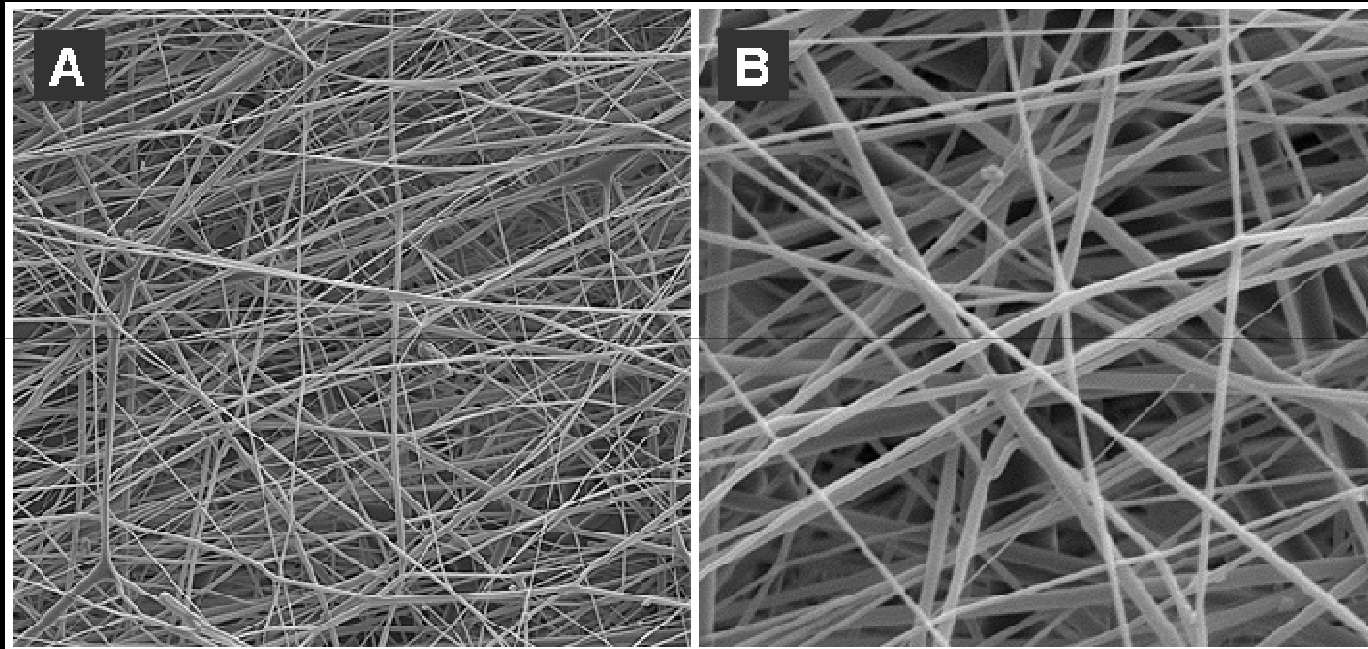
Electrospun fibrous mats in food-related separation applications

Veleirinho, B. and Lopes-da-Silva, J.A. (2009)
Process Biochemistry, 44, pp. 353-356.



- fibrous mats composed of randomly oriented submicron-size fibers can be prepared by electrospinning and successfully used as nonwoven filters in fruit juice clarification

Electrospun fibrous mats in food-related separation applications



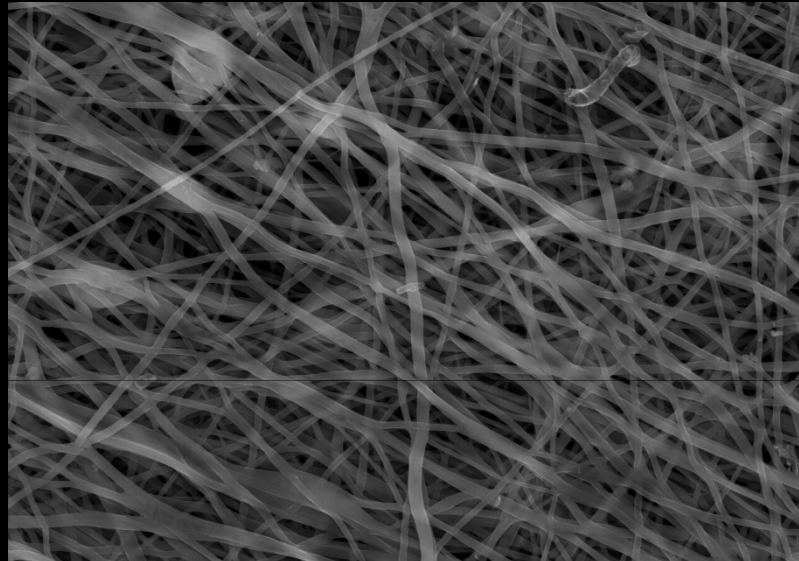
- SEM images showing the morphology of the PET electrospun membrane (A) x 500; (B) x 1500.

Electrospun fibrous mats in food-related separation applications

Open area for further developments:

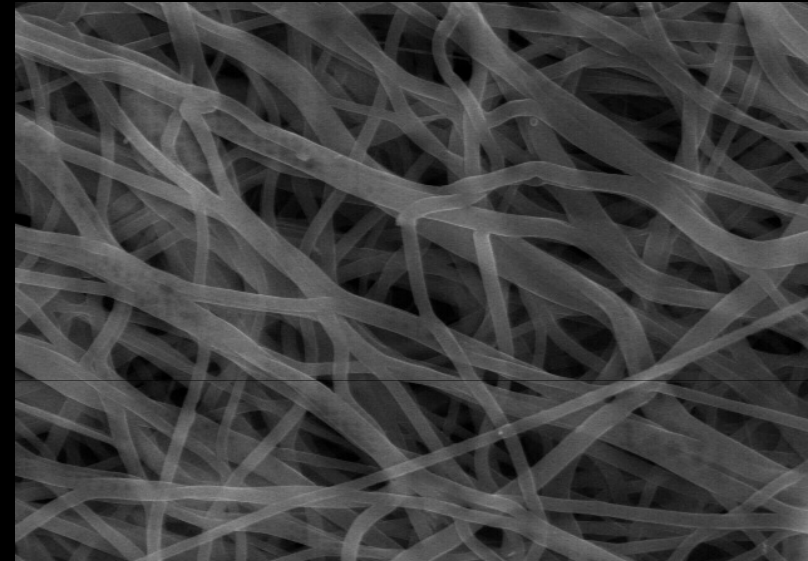
- multifunctional electrospun fiber membranes able to operate not only on separation mechanisms, but also showing additional functionalities.
 - Antimicrobial nanofibrous membranes to improve the microbial safety of beverages;
 - Development of nanofibrous electrospun mats to be used as separation membranes and improved carriers for enzymes enabling concomitant filtration and catalytic functionalities.

Encapsulation of β -galactosidase in PVA nanofibrous membranes



SU-70 15.0kV 15.4mm x5.00k SE(M)

10.0um



SU-70 15.0kV 15.4mm x10.0k SE(M)

5.00um

SEM images of electrospun PVA/ β -Galactosidase nanofibers

Encapsulation of β -galactosidase in PVA nanofibrous membranes

Enzyme loading efficiency

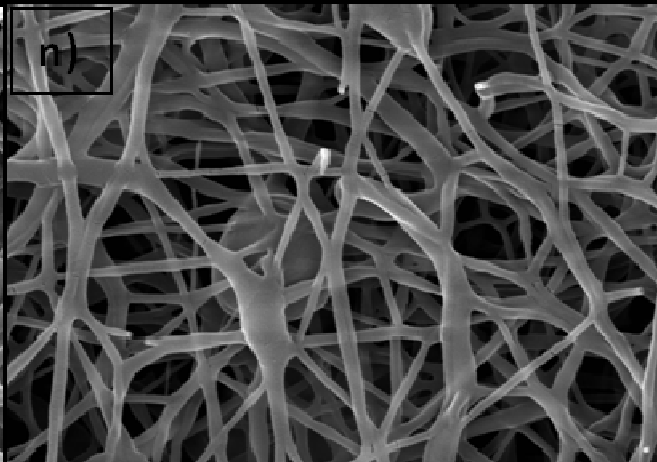
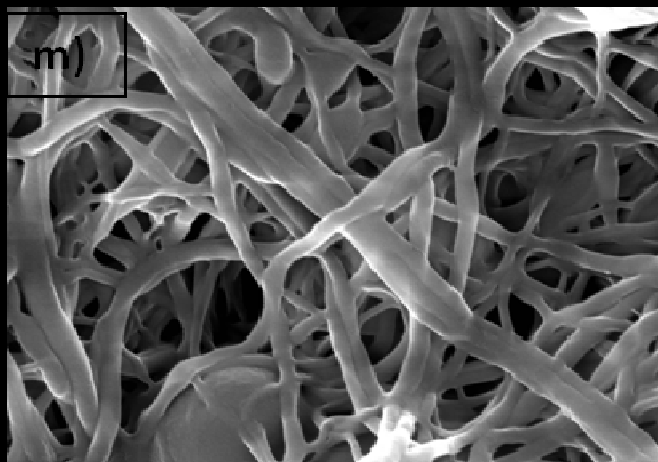
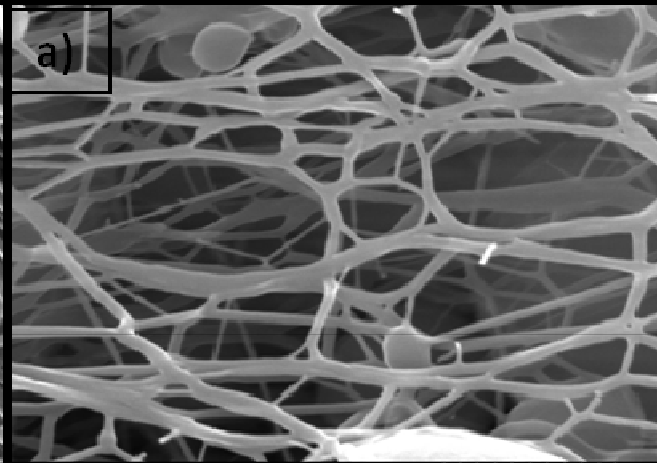
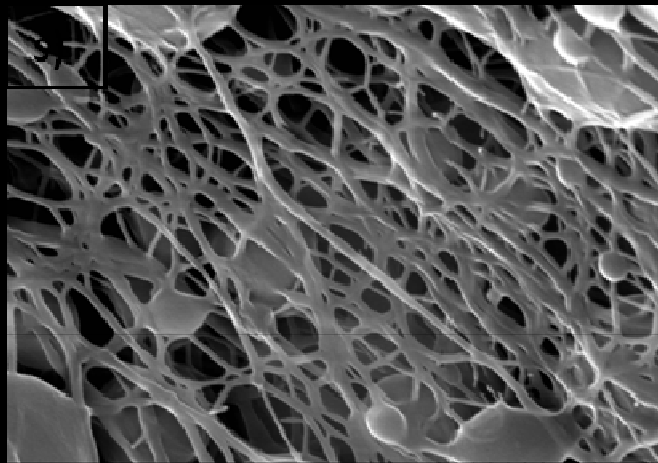
Time of immersion (min)	[Protein] (mg/mL)	Protein in Solution (mg)	% immobilization
30	0,08	0,412	83,5
60	0,08	0,417	83,3
220	0,084	0,421	83,1
280	0,08	0,415	83,4

Encapsulation of β -galactosidase in PVA nanofibrous membranes

Activity of free and immobilized β -Galactosidase

	Free β -Gal			Immobilized β -Gal		
Days	Activity (mmol min ⁻¹)	Specific Activity (mmol min ⁻¹ mg ⁻¹)	Retained activity (%)	Activity (mmol min ⁻¹)	Specific Activity (mmol min ⁻¹ mg ⁻¹)	Retained activity (%)
0	7.36×10^{-5}	9.81×10^{-4}	N.A.	N.A.	N.A.	N.A.
5	6.24×10^{-5}	8.33×10^{-4}	84.9	7.84×10^{-6}	3.19×10^{-5}	3.25
7	6.94×10^{-5}	9.26×10^{-4}	94.3	6.31×10^{-6}	2.51×10^{-5}	2.55
11	6.49×10^{-5}	8.65×10^{-4}	88.2	4.32×10^{-6}	1.78×10^{-5}	1.81

Encapsulation of trypsin in PCL nanofibers by emulsion electrospinning



Acknowledgements

- My students,
Bia Veleirinho, Pedro Ferreira, A. Rita Teles,
Diogo Silva, Nuno Almeida and Susana Pinto
- Financial support made possible our work
 - FEDER, through “Programa Operacional Factores de Competitividade” – COMPETE, and FCT – Fundação para a Ciência e a Tecnologia - project NanoBioCats (PTDC/CTM-POL/112289/2009)
 - QOPNA research unit (project PEst-C/QUI/UI0062/2011)

Aveiro Portugal

